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WORLD INTELLECTUAL PROPERTY ORGANIZATION  
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INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

<b>(51) International Patent Classification <sup>7</sup> :</b> C12N 15/10, 15/11, 5/10, 7/04, A61K 39/21, C07B 61/00, G06F 19/00, B01J 19/00 // C12N 15/12, 15/55, 15/49		<b>A3</b>	<b>(11) International Publication Number:</b> WO 00/18906 <b>(43) International Publication Date:</b> 6 April 2000 (06.04.00)
<b>(21) International Application Number:</b> PCT/US99/22588 <b>(22) International Filing Date:</b> 28 September 1999 (28.09.99) <b>(30) Priority Data:</b> 60/102,362 29 September 1998 (29.09.98) US 60/117,729 29 January 1999 (29.01.99) US 60/118,813 5 February 1999 (05.02.99) US 60/141,049 24 June 1999 (24.06.99) US <b>(71) Applicant (for all designated States except US):</b> MAXYGEN, INC. [US/US]; 515 Galveston Drive, Redwood City, CA 94063 (US). <b>(72) Inventors; and</b> <b>(75) Inventors/Applicants (for US only):</b> PATTEN, Phillip, A. [US/US]; Apartment 506, 2680 Fayette Drive, Mountain View, CA 94040 (US). LIU, Lu [US/US]; 519 Skiff Circle, Redwood City, CA 94056 (US). STEMMER, Willem, P., C. [NL/US]; 108 Kathy Court, Los Gatos, CA 95030 (US). <b>(74) Agents:</b> QUINE, Jonathan, Alan; Law Offices of Jonathan Alan Quine, P.O. Box 458, Alameda, CA 94501 (US) et al.		<b>(81) Designated States:</b> AE, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CU, CZ, DE, DK, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, UA, UG, US, UZ, VN, YU, ZA, ZW, ARIPO patent (GH, GM, KE, LS, MW, SD, SL, SZ, TZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG).  <b>Published</b> <i>With international search report.</i>  <b>(88) Date of publication of the international search report:</b> 26 October 2000 (26.10.00)	
<b>(54) Title:</b> SHUFFLING OF CODON ALTERED GENES			
<b>(57) Abstract</b>  Methods of recombining codon-altered libraries of nucleic acids are provided. The nucleic acids can include conservative or non-conservative modifications of coding sequences, in addition to codon alterations, as compared with wild-type sequences. In addition to making new proteins, methods of generation vectors with reduced rates of reversion to wild-type and attenuated viruses are also provided.			



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<p>(21) International Application Number: <b>PCT/US99/22588</b></p> <p>(22) International Filing Date: 28 September 1999 (28.09.99)</p> <p>(30) Priority Data:</p> <table border="0"> <tr> <td>60/102,362</td> <td>29 September 1998 (29.09.98)</td> <td>US</td> </tr> <tr> <td>60/117,729</td> <td>29 January 1999 (29.01.99)</td> <td>US</td> </tr> <tr> <td>60/118,813</td> <td>5 February 1999 (05.02.99)</td> <td>US</td> </tr> <tr> <td>60/141,049</td> <td>24 June 1999 (24.06.99)</td> <td>US</td> </tr> </table> <p>(71) Applicant (for all designated States except US): MAXYGEN, INC. [US/US]; 515 Galveston Drive, Redwood City, CA 94063 (US).</p> <p>(72) Inventors; and</p> <p>(75) Inventors/Applicants (for US only): PATTEN, Phillip, A. [US/US]; Apartment 506, 2680 Fayette Drive, Mountain View, CA 94040 (US). LIU, Lu [US/US]; 519 Skiff Circle, Redwood City, CA 94056 (US). STEMMER, Willem, P., C. [NL/US]; 108 Kathy Court, Los Gatos, CA 95030 (US).</p> <p>(74) Agents: QUINE, Jonathan, Alan; Law Offices of Jonathan Alan Quine, P.O. Box 458, Alameda, CA 94501 (US) et al.</p>	60/102,362	29 September 1998 (29.09.98)	US	60/117,729	29 January 1999 (29.01.99)	US	60/118,813	5 February 1999 (05.02.99)	US	60/141,049	24 June 1999 (24.06.99)	US	<p>(81) Designated States: AE, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CU, CZ, DE, DK, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, UA, UG, US, UZ, VN, YU, ZA, ZW, ARIPO patent (GH, GM, KE, LS, MW, SD, SL, SZ, TZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG).</p> <p><b>Published</b> <i>With international search report.</i></p> <p>(88) Date of publication of the international search report: 26 October 2000 (26.10.00)</p>
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# INTERNATIONAL SEARCH REPORT

Inter. Appl. No.

PCT/US 99/22588

## A. CLASSIFICATION OF SUBJECT MATTER

IPC 7 C12N15/10 C12N15/11 C12N5/10 C12N7/04 A61K39/21  
C07B61/00 G06F19/00 B01J19/00 //C12N15/12, C12N15/55,  
C12N15/49

According to International Patent Classification (IPC) or to both national classification and IPC

## B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 7 C12N C07K A61K C07B G06F B01J

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

EPO-Internal, BIOSIS, WPI Data

## C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	WO 97 35966 A (MAXYGEN INC ;MINSHULL JEREMY (US); STEMMER WILLEM P C (US)) 2 October 1997 (1997-10-02) abstract page 25, line 1 - line 29	1-49
X	WO 98 13485 A (ES HELMUTH H G VAN ;MAXYGEN INC (US); STEMMER WILLEM P C (US)) 2 April 1998 (1998-04-02) abstract page 9, line 28 -page 10, line 13 page 46, line 31 -page 47, line 34	1-49
A	----- -/--	50,51

☒ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

\* Special categories of cited documents :

- "A" document defining the general state of the art which is not considered to be of particular relevance
- "E" earlier document but published on or after the international filing date
- "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)
- "O" document referring to an oral disclosure, use, exhibition or other means
- "P" document published prior to the international filing date but later than the priority date claimed

- "T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
- "X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
- "Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.
- "&" document member of the same patent family

Date of the actual completion of the international search

18 July 2000

Date of mailing of the international search report

28.07.00

Name and mailing address of the ISA

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Authorized officer

Galli, I

# INTERNATIONAL SEARCH REPORT

International Application No

PCT/US 99/22588

## C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	S RAYNER ET AL: "MerMade: An oligodeoxyribonucleotide synthesizer for high throughput oligonucleotide production in dual 96-well plates" PCR METHODS AND APPLICATIONS,US,COLD SPRING HARBOR, NY, vol. 8, no. 7, 1 July 1998 (1998-07-01), pages 741-747, XP002089330 ISSN: 1054-9803 the whole document	50,51
X	WO 92 06176 A (IXSYS INC) 16 April 1992 (1992-04-16) abstract claims 22-30	36-42, 46,47,49
Y		43-45,48
Y	PATTEN P A ET AL: "APPLICATIONS OF DNA SHUFFLING TO PHARMACEUTICALS AND VACCINES" CURRENT OPINION IN BIOTECHNOLOGY,GB,LONDON, vol. 8, 1997, pages 724-733, XP002916609 ISSN: 0958-1669 the whole document	43-45,48
A	KIM C H ET AL: "Codon optimization for high-level expression of human erythropoietin (EPO) in mammalian cells" GENE: AN INTERNATIONAL JOURNAL ON GENES AND GENOMES,GB,ELSEVIER SCIENCE PUBLISHERS, BARKING, vol. 199, no. 1-2, 15 October 1997 (1997-10-15), pages 293-301, XP004126394 ISSN: 0378-1119 the whole document	17,45
A	AKASHI H: "Codon bias evolution in Drosophila. Population genetics of mutation-selection drift" GENE: AN INTERNATIONAL JOURNAL ON GENES AND GENOMES,GB,ELSEVIER SCIENCE PUBLISHERS, BARKING, vol. 205, no. 1-2, 31 December 1997 (1997-12-31), pages 269-278, XP004102913 ISSN: 0378-1119 the whole document	1-49

# INTERNATIONAL SEARCH REPORT

International application No.  
PCT/US 99/22588

## B x I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This International Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☐ Claims Nos.:  
because they relate to subject matter not required to be searched by this Authority, namely:
  
2. ☒ Claims Nos.:  
because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically:  
  
Claim 42 is dependent on claim 25. Since it relates to a library, it should be dependent on any of claims 38-41. Claim 43 is dependent on claim 41, but since it relates to a method, it should be dependent on claim 36.
  
3. ☐ Claims Nos.:  
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

## Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

see additional sheet

As a result of the prior review under R. 40.2(e) PCT,  
no additional fees are to be refunded.

1. ☐ As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.
  
2. ☐ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
  
3. ☒ As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:  
  
Invention 1 (claims 1-49) and invention 8 (claims 50-51)
  
4. ☐ No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

- ☒ The additional search fees were accompanied by the applicant's protest.
- ☐ No protest accompanied the payment of additional search fees.

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

This International Searching Authority found multiple (groups of) inventions in this international application, as follows:

1. Claims: (1-49) - partly, where applicable

A method of making codon-altered nucleic acids, comprising (i) providing a first nucleic acid sequence encoding a first polypeptide, (ii) providing a plurality of codon-altered nucleic acid sequences, each of which encode the first polypeptide or a modified form thereof, and (iii) recombining the plurality of codon-altered nucleic acid sequences to produce a target codon-altered nucleic acid, which encodes a second polypeptide.

Corresponding methods to make a library, said library per se, recombinant vectors carrying said codon-altered sequences.

Said methods and materials applied to sequences encoding erythropoietin.

2. Claims: (1-49) - partly, where applicable

Idem as subject-matter 1, but limited to sequences encoding G-CSF.

3. Claims: (1-49) - partly, where applicable

Idem as subject-matter 1, but limited to sequences encoding a viral envelope protein.

4. Claims: (1-49) - partly, where applicable

Idem as subject-matter 1, but limited to sequences encoding a cytokine.

5. Claims: (1-49) - partly, where applicable

Idem as subject-matter 1, but limited to sequences encoding a phosphatase.

6. Claims: (1-49) - partly, where applicable

Idem as subject-matter 1, but limited to sequences which, once recombined with a portion of a viral genome, produce an attenuated virus.

7. Claims: (1-49) - partly, where applicable

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

Idem as subject-matter 1, but wherein the target codon-altered nucleic acid is recombined with a portion of a viral genome to produce a viral vector that requires trans-complementation for replication and has a reduced rate of reversion to a replicative form.

8. Claims: (50,51) - completely

An integrated system comprising a computer or computer readable medium comprising a database having at least two artificial homologous codon-altered nucleic acids strings and a user interface allowing a user to selectively view one or more sequence strings in the database.



# INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No

PCT/US 99/22588

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
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		US 5770434 A	23-06-1998



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<b>(54) Title: SHUFFLING OF CODON ALTERED GENES</b> <b>(54) Titre: REARRANGEMENT DE GENES MODIFIES PAR CODON</b>  <b>(57) Abstract</b> <p>Methods of recombining codon-altered libraries of nucleic acids are provided. The nucleic acids can include conservative or non-conservative modifications of coding sequences, in addition to codon alterations, as compared with wild-type sequences. In addition to making new proteins, methods of generation vectors with reduced rates of reversion to wild-type and attenuated viruses are also provided.</p> <b>(57) Abrégé</b> <p>La présente invention concerne des techniques de recombinaison de banques, modifiées par codon, d'acides nucléiques. En plus des modifications par codon, ces acides nucléiques peuvent inclure des modifications conservatrices ou non conservatrices des séquences de codage, par opposition à des séquences de type sauvage. L'invention concerne l'obtention, outre de nouvelles protéines, de vecteurs caractérisés par des taux réduits de retransformation en virus de type sauvage et atténué.</p>		

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**Description**

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**CROSS REFERENCE TO RELATED APPLICATIONS**

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This application is a non-provisional filing of USSN 60/102,362,

- 5 "SHUFFLING OF CODON ALTERED GENES," Attorney Docket No. 02-028500, by Patten and Stemmer, filed 09/29/98, and 60/117,729, "SHUFFLING OF CODON ALTERED GENES," Attorney Docket No. 02-028510, by Patten and Stemmer, filed January 29, 1999.
- 15 The application is also related to USSN 60/118,813 "OLIGONUCLEOTIDE MEDIATED NUCLEIC ACID RECOMBINATION," by Cramer et al., Attorney Docket Number 02-296,
- 10 filed February 5, 1999; and USSN 60/141,049 "OLIGONUCLEOTIDE MEDIATED NUCLEIC ACID RECOMBINATION," by Cramer et al., Attorney Docket Number 02-296-
- 20 1, filed June 24, 1999.

**BACKGROUND**

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The genetic code is highly degenerate. Every DNA/RNA triplet (codon)

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- encoding an amino acid can typically be altered, with the exception of ATG/AUG (coding for methionine) and TGG/UGG (coding for Tryptophan), without altering the sequence of the protein encoded by the corresponding nucleic acid sequence. Roughly, on average (the
- 30 distribution of amino acids varies from protein to protein), each coding triplet can be substituted about 3 different ways, since there are 61 codons encoding 20 amino acids (there
- 20 are 3 additional triplets encoding stop codons, for a total of 64 codons encoding 20 amino acids). This represents a possible sequence diversity of approximately  $3^n$  possible sequences
- 35 which encode a given protein, where  $n$  is the length of the protein in amino acids. As can easily be seen, for proteins of even modest length, the number of possible nucleic acids which can encode the protein exceeds the number of physical particles in the universe (estimated at
- 40 25 about  $10^{80}$  particles).

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- This tremendous potential coding sequence space for individual proteins has interesting evolutionary implications. For example, hypermutable viruses such as HIVs and other retroviruses typically stay one step ahead of the host immune system by accumulating
- 50 non-random mutations based, in part, upon the particular codons used to encode recognition molecules, e.g., in the envelope portion of the virus. The mutations are non-random because viruses are selected for the ability to mutate to forms which are not quickly recognized by the

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5 host immune system. A consequence of this is that viruses are selected to have a non-random set of codons encoding, e.g., envelope proteins, allowing the viruses to shift forms rapidly by making, e.g., specific point mutations to generate specific alterations in protein structure.

10 Codon use is also non-random within species. By preferentially making a  
5 subset of all possible t-RNAs, cells may conserve energy, and can optimize, or even regulate, the efficiency of cellular translation systems. This fact has long been recognized empirically, often allowing investigators initially to determine the reading frame of a given nucleic acid  
15 sequence simply by consideration of the codons resulting from different potential reading frames. One consequence of this "species codon bias" is that proteins within a species have a  
10 limited set of possible mutations that can arise as a consequence of, e.g., point mutation. This limits the possible evolution rate of proteins.

In addition to the diversity of nucleic acid coding sequences which encode any given protein, it is now clear that protein sequences are, themselves, quite degenerate. Often,  
25 many of the amino acid residues constituting a protein may be substituted for structurally  
15 similar amino acid units without significantly changing the tertiary structure of the protein. Thus, it may be difficult to determine which residues to modify or to improve desirable properties of a protein.

30 For proteins which are commercially valuable, it would be desirable to be able to gain access to a mutational spectrum which is different than that of the native protein. The  
20 present invention provides this, and many other features, that will be apparent upon complete review of the following.

#### SUMMARY OF THE INVENTION

The present invention provides methods of accessing a completely different  
40 mutational spectrum for a selected protein than is available in the naturally occurring nucleic  
25 acid encoding the protein. This increases the type and rate of forced evolution for the selected protein, allowing for rapid improvement of any detectable characteristic of the protein. In the methods, nucleic acids are synthesized with altered codon usage, and/or  
45 which encode one or several amino acid residue changes as compared to the selected protein, where the amino acid and codon usage changes can be conservative or non-conservative.  
30 The resulting codon/amino acid modified nucleic acid(s) are recombined using DNA shuffling techniques with either the native nucleic acid, or with each other (or both), typically

5 using recursive shuffling methods. The nucleic acids or the encoded protein are then screened for a desirable property.

Thus, the invention provides methods of making codon altered nucleic acids.

10 In the methods, a first nucleic acid sequence encoding a first polypeptide sequence is selected. A plurality of codon altered nucleic acid sequences, each of which encode the first polypeptide, or a modified form thereof, are then selected (e.g., a library of codon altered  
15 nucleic acids can be selected in a biological assay which recognizes library components or activities), and the plurality of codon altered nucleic acid sequences is recombined to produce a target codon altered nucleic acid encoding a second protein. The target codon altered  
20 nucleic acid is then screened for a detectable functional or structural property, optionally including comparison to the properties of the first polypeptide. The goal of such screening is to identify a polypeptide that has a structural or functional property equivalent or superior to the first polypeptide. A nucleic acid encoding such a polypeptide can be used in essentially  
25 any procedure desired, including introducing the target codon altered nucleic acid into a cell, vector, virus, attenuated virus (e.g., as a component of a vaccine or immunogenic composition), transgenic organism, or the like.

30 Kits and compositions for practicing the methods are also provided, including one or more of: cell recombination mixtures and substrates (e.g., nucleic acids with altered codon usage), containers, instructional material for practicing the methods, or the like.

#### 20 BRIEF DESCRIPTION OF THE FIGURES

35 Figure 1 is a nucleic acid/amino acid sequence of a part of the monkey EPO gene, which is similar to the human EPO gene.

Figure 2 shows an example of a codon altered EPO nucleic acid sequence.

Figure 3 shows an alignment of naturally occurring EPOs.

40 25 Figure 4 is a schematic of the human EPO wobble sequence space.

Figure 5 is a schematic of Mammalian EPO Family-Wobble Sequence Space.

45 Figure 6 is a sequence alignment of G-CSF homologs, with species information.

30 Figure 7 is a sequence alignment of G-CSF homologs, with differences broken out.



Figure 8 is a sequence alignment showing the hydrophobic core residues of human G-CSF (blacked out).

Figure 9 is a schematic showing the shuffling strategy for G-CSF.

Figure 10 is a list of oligos used to make a codon altered alkaline phosphatase.

Figure 11 is a map of oligos used to make a codon altered alkaline phosphatase.

Figure 12 is a schematic of vaccination with evolution defective viruses.

Figure 13 is a schematic of different mutations that result from different codon types for ser, arg, and leu.

Figure 14 is a schematic of vaccination with evolution defective viruses.

Figure 15 is a schematic of vaccination with evolution defective viruses showing sophisticated versus non-sophisticated "mutant clouds."

Figure 16, panels A-C show results of single mutations of different codons for scr, arg, and leu.

Fig. 17 is a schematic of protein evolution with expanded mutation spectra.

Fig. 18, panels A-D show codon altered forms of Env.

Fig. 19 is a list of oligos in one application for synthesis of HIV Env.

#### DEFINITIONS

Unless clearly indicated to the contrary, the following definitions supplement definitions of terms known in the art.

As used herein, a "recombinant" nucleic acid is a nucleic acid produced by recombination between two or more nucleic acids, or any nucleic acid made by an *in vitro* or artificial process. The term "recombinant" when used with reference to a cell indicates that the cell comprises (and optionally replicates) a heterologous nucleic acid, or expresses a peptide or protein encoded by a heterologous nucleic acid. Recombinant cells can contain genes that are not found within the native (non-recombinant) wild-type form of the cell. Recombinant cells can also contain genes found in the native form of the cell where the genes are modified and re-introduced into the cell by artificial means. The term also encompasses cells that contain a nucleic acid endogenous to the cell that has been artificially modified without removing the nucleic acid from the cell; such modifications include those obtained by gene replacement, site-specific mutation, chimeraplasty, and related techniques.

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A "codon altered" nucleic acid is a first nucleic acid that encodes a first polypeptide similar or identical to a naturally occurring polypeptide encoded by a naturally occurring nucleic acid, where the first nucleic acid utilizes a plurality of codons to encode the first polypeptide, which differ from the codons of the naturally occurring nucleic acid that encode the naturally occurring polypeptide.

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A "nucleic acid sequence" refers to either a nucleic acid (e.g., RNA, DNA or modified form thereof, in isolated, recombinant or native form) or to a representation of the nucleic acid such as a sequence of letters indicating the primary structure (sequence) of the nucleic acid.

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A "polypeptide sequence" refers to either a polypeptide (or modified form thereof, in isolated, recombinant or native form) or to a representation of the polypeptide such as a sequence of letters or other character string information indicating the primary structure (amino acid sequence) of the polypeptide.

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A "modified form" of a reference polypeptide is a target polypeptide which has a similar, but not identical, sequence to the reference polypeptide. The sequence of the target polypeptide can differ from the reference polypeptide by conservative or non-conservative substitutions of the reference polypeptide sequence. As noted in more detail, *supra*, different nucleic acids encoding different target polypeptides having different non-conservative substitutions relative to the reference polypeptide can be recombined to produce a recombined nucleic acid encoding a target polypeptide more similar to the reference polypeptide.

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A "plurality of forms" of a selected nucleic acid refers to a plurality of homologs of the nucleic acid. The homologs can be from naturally occurring homologs (e.g., two or more homologous genes, or derivatives thereof) or by artificial synthesis of one or more nucleic acids having related sequences, or by modification of one or more nucleic acid to produce related nucleic acids. Nucleic acids are homologous when they are derived, naturally or artificially, from a common ancestor sequence. During natural evolution, this occurs when two or more descendent sequences diverge from a parent sequence over time, i.e., due to mutation and natural selection. Under artificial conditions, divergence occurs, e.g., in one of two ways. First, a given sequence can be artificially recombined with another sequence, as occurs, e.g., during typical cloning, to produce a descendent nucleic acid.

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5 Alternatively, a nucleic acid can be synthesized *de novo*, by synthesizing a nucleic acid which varies in sequence from a given parental nucleic acid sequence.

10 When there is no explicit knowledge about the ancestry of two nucleic acids, homology is typically inferred by sequence comparison between two sequences. Where two  
5 nucleic acid sequences show sequence similarity it is inferred that the two nucleic acids share a common ancestor. The precise level of sequence similarity required to establish homology varies in the art depending on a variety of factors. For purposes of this disclosure, two  
15 sequences are considered homologous where they share sufficient sequence identity to allow recombination to occur between two nucleic acid molecules, or when codon changes can be  
10 made which would result in two or more nucleic acids having the ability to recombine. Typically, nucleic acids require regions of close similarity spaced roughly the same distance  
20 apart to permit recombination to occur.

25 The terms "identical" or percent "identity," in the context of two or more nucleic acid or polypeptide sequences, refer to two or more sequences or subsequences that  
15 are the same or have a specified percentage of amino acid residues or nucleotides that are the same, when compared and aligned for maximum correspondence, as measured using one of the sequence comparison algorithms described below (or other algorithms available to  
30 persons of skill) or by visual inspection.

35 The phrase "substantially identical," in the context of two nucleic acids or polypeptides refers to two or more sequences or subsequences that have at least about 40%,  
20 50%, 60%, or preferably about 70% or 80% or more, or most preferably 90-95% nucleotide or amino acid residue identity, when compared and aligned for maximum correspondence, as measured using one of the following sequence comparison algorithms or by visual inspection.  
Such "substantially identical" sequences are typically considered to be homologous.  
40 25 Preferably, the "substantial identity" exists over a region of the sequences that is at least about 50 residues in length, more preferably over a region of at least about 100 residues, and most preferably the sequences are substantially identical over at least about 150 residues, or  
45 over the full length of the two sequences to be compared.

50 For sequence comparison and homology determination, typically one sequence acts as a reference sequence to which test sequences are compared. When using a sequence comparison algorithm, test and reference sequences are input into a computer,

subsequence coordinates are designated, if necessary, and sequence algorithm program parameters are designated. The sequence comparison algorithm then calculates the percent sequence identity for the test sequence(s) relative to the reference sequence, based on the designated program parameters.

- Optimal alignment of sequences for comparison can be conducted, e.g., by the local homology algorithm of Smith & Waterman, *Adv. Appl. Math.* 2:482 (1981), by the homology alignment algorithm of Needleman & Wunsch, *J. Mol. Biol.* 48:443 (1970), by the search for similarity method of Pearson & Lipman, *Proc. Nat'l. Acad. Sci. USA* 85:2444 (1988), by computerized implementations of these algorithms (GAP, BESTFIT, FASTA, and TFASTA in the Wisconsin Genetics Software Package, Genetics Computer Group, 575 Science Dr., Madison, WI), or by visual inspection (*see generally*, Ausubel *et al.*, *infra*).

- One example of algorithm that is suitable for determining percent sequence identity and sequence similarity is the BLAST algorithm, which is described in Altschul *et al.*, *J. Mol. Biol.* 215:403-410 (1990). Software for performing BLAST analyses is publicly available through the National Center for Biotechnology Information (<http://www.ncbi.nlm.nih.gov/>). This algorithm involves first identifying high scoring sequence pairs (HSPs) by identifying short words of length W in the query sequence, which either match or satisfy some positive-valued threshold score T when aligned with a word of the same length in a database sequence. T is referred to as the neighborhood word score threshold (Altschul *et al.*, *supra*). These initial neighborhood word hits act as seeds for initiating searches to find longer HSPs containing them. The word hits are then extended in both directions along each sequence for as far as the cumulative alignment score can be increased. Cumulative scores are calculated using, for nucleotide sequences, the parameters M (reward score for a pair of matching residues; always > 0) and N (penalty score for mismatching residues; always < 0). For amino acid sequences, a scoring matrix is used to calculate the cumulative score. Extension of the word hits in each direction are halted when: the cumulative alignment score falls off by the quantity X from its maximum achieved value; the cumulative score goes to zero or below, due to the accumulation of one or more negative-scoring residue alignments; or the end of either sequence is reached. The BLAST algorithm parameters W, T, and X determine the sensitivity and speed of the alignment. The BLASTN program (for nucleotide sequences) uses as defaults a wordlength (W) of 11, an expectation

(E) of 10, a cutoff of 100, M=5, N=-4, and a comparison of both strands. For amino acid sequences, the BLASTP program uses as defaults a wordlength (W) of 3, an expectation (E) of 10, and the BLOSUM62 scoring matrix (*see* Henikoff & Henikoff (1989) *Proc. Natl. Acad. Sci. USA* 89:10915).

In addition to calculating percent sequence identity, the BLAST algorithm also performs a statistical analysis of the similarity between two sequences (*see, e.g.,* Karlin & Altschul (1993) *Proc. Nat'l. Acad. Sci. USA* 90:5873-5787). One measure of similarity provided by the BLAST algorithm is the smallest sum probability (P(N)), which provides an indication of the probability by which a match between two nucleotide or amino acid sequences would occur by chance. For example, a nucleic acid is considered similar to a reference sequence if the smallest sum probability in a comparison of the test nucleic acid to the reference nucleic acid is less than about 0.1, more preferably less than about 0.01, and most preferably less than about 0.001.

Another indication that two nucleic acid sequences are substantially identical/homologous is that the two molecules hybridize to each other under stringent conditions. The phrase "hybridizing specifically to," refers to the binding, duplexing, or hybridizing of a molecule only to a particular nucleotide sequence under stringent conditions, including when that sequence is present in a complex mixture (*e.g.,* total cellular) DNA or RNA. "Bind(s) substantially" refers to complementary hybridization between a probe nucleic acid and a target nucleic acid and embraces minor mismatches that can be accommodated by reducing the stringency of the hybridization media to achieve the desired detection of the target polynucleotide sequence.

"Stringent hybridization conditions" and "stringent hybridization wash conditions" in the context of nucleic acid hybridization experiments such as Southern and northern hybridizations are sequence dependent, and are different under different environmental parameters. Longer sequences and sequences with higher G:C content remain hybridized at higher temperatures (or at lower salt). An extensive guide to the hybridization of nucleic acids is found in Tijssen (1993) *Laboratory Techniques in Biochemistry and Molecular Biology--Hybridization with Nucleic Acid Probes* part I chapter 2 "Overview of principles of hybridization and the strategy of nucleic acid probe assays," Elsevier, New York.

5 Generally, highly stringent hybridization and wash conditions are selected to  
be about 5 °C lower than the thermal melting point ( $T_m$ ) for the specific sequence at a defined  
10 ionic strength and pH. Typically, under "stringent conditions" a probe will hybridize to its  
target subsequence, but not to unrelated (non-homologous) sequences.

5 The  $T_m$  is the temperature (under defined ionic strength and pH) at which 50%  
of the target sequence hybridizes to a perfectly matched probe. Very stringent conditions are  
15 selected to be equal to the  $T_m$  for a particular probe. An example of stringent hybridization  
conditions for hybridization of complementary nucleic acids which have more than 100  
complementary residues on a filter in a Southern or northern blot is 50% formamide with 1  
20 mg of heparin at 42 °C, with the hybridization being carried out overnight. An example of  
highly stringent wash conditions is 0.15M NaCl at 72 °C for about 15 minutes. An example  
of stringent wash conditions is a 0.2x SSC wash at 65 °C for 15 minutes (*see*, Sambrook,  
25 *infra.*, for a description of SSC buffer). Often, a high stringency wash is preceded by a low  
stringency wash to remove background probe signal. An example medium stringency wash  
15 for a duplex of, *e.g.*, more than 100 nucleotides, is 1x SSC at 45 °C for 15 minutes. An  
example low stringency wash for a duplex of, *e.g.*, more than 100 nucleotides, is 4-6x SSC at  
30 40 °C for 15 minutes. For short probes (*e.g.*, about 10 to 50 nucleotides), stringent conditions  
typically involve salt concentrations of less than about 1.0 M Na ion, typically about 0.01 to  
1.0 M Na ion concentration (or other salts) at pH 7.0 to 8.3, and the temperature is typically  
20 at least about 40 °C. Stringent conditions can also be achieved with the addition of  
destabilizing agents such as formamide. In general, a signal to noise ratio of 2x (or higher)  
35 than that observed for an unrelated probe in the particular hybridization assay indicates  
detection of a specific hybridization. If the signal to noise ratio is less than 2x binding of an  
unrelated probe (*e.g.*, a nucleic acid encoding a non-homologous protein), the nucleic acids at  
40 25 issue do not hybridize under stringent conditions. Similarly, if the signal to noise ratio is less  
than 25% as high as that observed for a perfectly matched probe under stringent conditions,  
the nucleic acids do not "hybridize under stringent conditions" as that term is used herein.  
This does not apply to highly stringent conditions, as the stringency can theoretically be  
45 increased until only a perfectly matched probe will hybridize.

30 In one example hybridization procedure, a target nucleic acid to be probed is  
blotted onto a filter by any conventional method. An unrelated nucleic acid such as a plasmid

5 vector (assuming that the target nucleic acid has no homology with the target nucleic acid) is  
also blotted, in approximately equal amounts onto the filter. The filter is probed with a  
10 labeled probe complementary to the target nucleic acid. The experiment is repeated at  
gradually increasing stringency of hybridization and wash conditions until signal from the  
5 hybridization of the labeled probe to the complementary target is 10-100X as high as to the  
unrelated plasmid vector nucleic acid. Once these conditions are determined as described  
15 above, a test nucleic acid is probed under the same conditions as the target. If signal from the  
labeled probe is 25% as high or higher than the signal from binding of the probe to the target,  
the test nucleic acid "hybridizes under stringent conditions" to the probe. If the signal is less  
20 than 25% as high, the test nucleic acid does not hybridize under stringent conditions to the  
probe.

Nucleic acids which do not hybridize to each other under stringent conditions  
are still recognizable as variant forms of a nucleic acid when the polypeptides they encode  
25 are substantially identical. This occurs, *e.g.*, when a copy of a nucleic acid is created using  
the maximum codon degeneracy permitted by the genetic code. Such nucleic acids are not  
15 functionally equivalent, as described in detail herein, due to differences in mRNA folding,  
alterations of regulatory sequences and the like.

30 Another indication that two nucleic acid sequences or polypeptides are variant  
forms is that the polypeptide encoded by the first nucleic acid is immunologically cross  
20 reactive with the polypeptide encoded by the second nucleic acid, as tested by polyclonal  
antisera generated to the first polypeptide. Thus, a polypeptide is typically substantially  
35 identical to a second polypeptide, for example, where the two peptides differ only by  
conservative substitutions.

40 "Conservatively modified variations" of a particular polynucleotide sequence  
25 are those polynucleotide variations that encode identical or essentially identical amino acid  
sequences, or where the polynucleotide does not encode an amino acid sequence, which  
encode essentially identical sequences. Because of the degeneracy of the genetic code, a large  
45 number of functionally identical nucleic acids encode any given polypeptide. For instance,  
the codons CGU, CGC, CGA, CGG, AGA, and AGG all encode the amino acid arginine.  
30 Thus, at every position where an arginine is specified by a codon, the codon can be altered to  
any of the corresponding codons described without altering the encoded polypeptide. Such

5 nucleic acid variations are "silent variations," which are one species of "conservatively modified variations." Every polynucleotide sequence described herein which encodes a polypeptide also optionally describes every possible silent variation, except where otherwise  
10 noted. One of skill will recognize that each codon in a nucleic acid (except AUG, which is ordinarily the codon for methionine, and TGG, which is ordinarily the codon for tryptophan) can be modified to yield a peptide which is structurally identical.

15 Furthermore, one of skill will recognize that individual substitutions, deletions or additions which alter, add or delete a single amino acid or a small percentage of amino acids (typically less than 5%, more typically less than 1%) in an encoded sequence are  
20 "conservatively modified variations" where the alterations result in the substitution of an amino acid with a chemically similar amino acid. Conservative substitution tables providing functionally similar amino acids are well known in the art. The following five groups each contain amino acids that are conservative substitutions for one another:

25 Aliphatic: Glycine (G), Alanine (A), Valine (V), Leucine (L), Isoleucine (I);  
15 Aromatic: Phenylalanine (F), Tyrosine (Y), Tryptophan (W); Sulfur-containing: Methionine (M), Cysteine (C); Basic: Arginine (R), Lysine (K), Histidine (H); Acidic: Aspartic acid (D), Glutamic acid (E), Asparagine (N), Glutamine (Q). *See also*, Creighton (1984) *Proteins*,  
30 W.H. Freeman and Company. In addition, individual substitutions, deletions or additions which alter, add or delete a single amino acid or a small percentage of amino acids in an  
20 encoded sequence are also "conservatively modified variations." Sequences that differ by conservative variations are generally homologous.

35 The term "isolated", when applied to a nucleic acid or protein, denotes that the nucleic acid or protein is essentially free of other cellular or other components (e.g., library components) with which it is associated in the natural state.

40 25 The term "nucleic acid" refers to deoxyribonucleotides or ribonucleotides and polymers thereof in either single- or double-stranded form. Unless specifically limited, the term encompasses nucleic acids containing known analogues of natural nucleotides which  
45 have similar binding properties as the reference nucleic acid and are metabolized in a manner similar to naturally occurring nucleotides. Unless otherwise indicated, a particular nucleic  
30 acid sequence also implicitly encompasses conservatively modified variants thereof (e.g. degenerate codon substitutions) and complementary sequences and as well as the sequence



5 explicitly indicated. Specifically, degenerate codon substitutions may be achieved by  
generating sequences in which the third position of one or more selected (or all) codons is  
10 substituted with mixed-base and/or deoxyinosine residues (Batzner *et al.* (1991) *Nucleic Acid*  
*Res.* 19: 5081; Ohtsuka *et al.* (1985) *J. Biol. Chem.* 260: 2605-2608; Cassol *et al.* (1992) ;  
5 Rossolini *et al.* (1994) *Mol. Cell. Probes* 8: 91-98). The term nucleic acid is generic to the  
terms "gene", "DNA," "cDNA", "oligonucleotide," "RNA," "mRNA," and the like.

15 "Nucleic acid derived from a gene" refers to a nucleic acid for whose  
synthesis the gene, or a subsequence thereof, has ultimately served as a template. Thus, an  
mRNA, a cDNA reverse transcribed from an mRNA, an RNA transcribed from that cDNA, a  
10 DNA amplified from the cDNA, an RNA transcribed from the amplified DNA, *etc.*, are all  
20 derived from the gene and detection of such derived products is indicative of the presence  
and/or abundance of the original gene and/or gene transcript in a sample.

A nucleic acid is "operably linked" when it is placed into a functional  
25 relationship with another nucleic acid sequence. For instance, a promoter or enhancer is  
15 operably linked to a coding sequence if it increases the transcription of the coding sequence.

A "recombinant expression cassette" or simply an "expression cassette" is a  
30 nucleic acid construct, generated recombinantly or synthetically, with nucleic acid elements  
that are capable of effecting expression of a structural gene in hosts compatible with such  
sequences. Expression cassettes include at least promoters and optionally, transcription  
20 termination signals. Typically, the recombinant expression cassette includes a nucleic acid to  
35 be transcribed (*e.g.*, a nucleic acid encoding a desired polypeptide), and a promoter.  
Additional factors necessary or helpful in effecting expression may also be used as described  
herein. For example, an expression cassette can also include nucleotide sequences that  
40 encode a signal sequence that directs secretion of an expressed protein from the host cell.  
25 Transcription termination signals, enhancers, and other nucleic acid sequences that influence  
gene expression, can also be included in an expression cassette.

#### DETAILED DISCUSSION OF THE INVENTION

45 In the present invention, the sequence diversity of substrates for DNA  
shuffling procedures is increased by using codon-altered nucleic acids as templates and/or by  
30 using templates that encode proteins with conservative or non-conservative amino acid  
modifications as compared to a selected wild-type protein.

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These codon altered nucleic acids can be chemically synthesized (e.g., using standard artificial synthetic protocols, e.g., those typically used by commercial sources from which nucleic acids can be ordered), or can be made using any of a variety of methods herein of available to one of skill. For example, oligonucleotide fragments can be made which correspond to a codon altered nucleic acid which is desired using standard synthetic methods, followed by polymerase and/or ligase mediated oligonucleotide ligation/recombination protocols to generate full-length nucleic acids.

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The combination of codon usage modifications and coding modifications can be extensive enough to reduce or, under stringent conditions, even eliminate the hybridization of the codon-altered nucleic acids to a nucleic acid which naturally encodes the selected protein. This dramatically alters the mutations which result from possible single nucleotide mutations, providing access to greater diversity for DNA shuffling protocols.

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In addition, the recombination and selection of such nucleic acids during DNA shuffling procedures can result not only in access to a different set of possible mutations, but can also result in modified forms of transcriptional or translational regulation, alterations in nucleic acid localization, mRNA stability and the like. Furthermore, the modified hybridization properties of codon altered nucleic acids leads to alterations in the ability of the nucleic acids to hybridize with potential recombination partners, altering, and ultimately increasing, the available recombination diversity during shuffling.

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Furthermore, "family shuffling" using codon-altered substrates even further increases the possible sequence diversity of the starting materials for recombination. As currently practiced, family shuffling methods involve shuffling nucleic acids encoding sequence variants of a given protein (e.g., species or allele homologs). In the present methods, this procedure is modified by generating codon-altered versions of the sequence variants to access additional molecular diversity during recombination. Additional diversity is achieved by conservatively and non-conservatively modifying the starting nucleic acids to encode non-naturally occurring sequence variants. Family shuffling can be performed even using homologs of relatively low identity. In such cases, codons may be changed in one or more of the family members to increase the level of identity between the members, thereby increasing their ability to recombine using the methods of this invention.

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Gene shuffling and family shuffling provide two of the most powerful methods available for improving and "migrating" (gradually changing the type of reaction, substrate or activity of a selected protein such as an enzyme, or regulation or structure of an expressed component) the functions of proteins. In family shuffling, homologous sequences, e.g., from different species, chromosomal positions, or due to synthetic alteration, are recombined. In gene shuffling, a single sequence is mutated or otherwise altered and then recombined.

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The generation and screening of high quality shuffled libraries provides for DNA shuffling (or "directed evolution"). The availability of appropriate high-throughput analytical chemistry to screen the libraries permits integrated high-throughput shuffling and screening of the libraries to achieve a desired activity.

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In one significant embodiment, oligonucleotides for constructing codon-modified nucleic acids are designed in a computer ("in silico"). Predicted codon-modified recombinant nucleic acids can also be determined in silico, i.e., essentially as taught in Selifonov and Stemmer "METHODS FOR MAKING CHARACTER STRINGS, POLYNUCLEOTIDES & POLYPEPTIDES HAVING DESIRED CHARACTERISTICS" filed 02/05/1999, USSN 60/118854.

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Furthermore, rather than generating codon-modified nucleic acids as substrates for recombination, families of nucleic acids can be recombined simply by appropriate selection of the relevant oligonucleotides which are used in gene reconstruction methods to produce recombinant nucleic acids, i.e., by using codon-modified nucleic acid oligonucleotides as discussed herein in conjunction with family oligonucleotide-mediated shuffling methods, e.g., as taught in Crameri et al. "OLIGONUCLEOTIDE MEDIATED NUCLEIC ACID RECOMBINATION" filed February 5, 1999, USSN 60/118,813 and Crameri et al. "OLIGONUCLEOTIDE MEDIATED NUCLEIC ACID RECOMBINATION" filed June 24, 1999, USSN 60/141,049. The technique can be used to recombine homologous or even non-homologous nucleic acid sequences; in the context of the present invention, oligonucleotides corresponding to families of codon-modified nucleic acids are shuffled.

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The present invention provides significant advantages over previously used methods for optimization of genes. For example, DNA shuffling of codon modified nucleic acids can result in optimization of a desirable property even in the absence of a detailed

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5 understanding of the mechanism by which the particular property is mediated. In addition,  
entirely new properties can be obtained upon shuffling of codon modified DNAs, i.e.,  
10 shuffled DNAs can encode polypeptides or RNAs with properties entirely absent in the  
parental DNAs which are shuffled. Thus, by modifying the codon usage and/or encoded  
5 amino acids of the relevant gene or other nucleic acid, molecular diversity is accessed and  
sequences can be shuffled to obtain desired, including entirely new, properties.

15 In general, sequence recombination can be achieved in many different formats  
and permutations of formats, as described in further detail below.

The targets for modification vary in different applications, as does the  
10 property sought to be acquired or improved. Examples of candidate targets for acquisition of  
a property or improvement in a property include genes that encode proteins which have  
enzymatic or therapeutic or other commercially useful activities. A more extensive listing is  
found *supra*; however, even this list is not intended to be limiting, as essentially any nucleic  
25 acid can be codon modified and shuffled, using one or more of the processes herein.

15 Shuffling methods use at least two variant forms of a starting target (the  
variant forms can be nucleic acids, or representations thereof, e.g., as character strings in a  
computer program). The variant forms of candidate codon-altered substrates can show  
30 substantial sequence or secondary structural similarity with each other, but they should also  
differ in at least one and preferably at least two positions. The initial diversity between forms  
20 can be the result of natural variation, e.g., the different variant forms (homologs) are obtained  
from different individuals or strains of an organism, or constitute related sequences from the  
35 same organism (e.g., allelic variations), or constitute homologs from different organisms  
(interspecific variants), or constitute artificial homologs, e.g., codon-altered nucleic acids  
encoding the same or a similar protein. Any or all of these sequences can represent or  
40 25 include codon altered nucleic acids.

Initial diversity can also be induced, e.g., the variant forms can be generated  
by error-prone transcription, such as an error-prone PCR or use of a polymerase which lacks  
45 proof-reading activity (see, Liao (1990) *Gene* 88:107-111), of the first variant form, or, by  
replication of the first form in a mutator strain (mutator host cells are discussed in further  
30 detail below, and are generally well known). The initial diversity between substrates is  
greatly augmented in subsequent steps of recombination for library generation.

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A mutator strain can include any mutants in any organism impaired in the functions of mismatch repair. These include mutant gene products of mutS, mutT, mutH, mutL, ovrD, dcm, vsr, umuC, umuD, sbcB, recJ, etc. The impairment is achieved by genetic mutation, allelic replacement, selective inhibition by an added reagent such as a small compound or an expressed antisense RNA, or other techniques. Impairment can be of the genes noted, or of homologous genes in any organism. The properties or characteristics that can be acquired or improved vary widely, and, of course depend on the choice of substrate.

20

At least two variant forms of a nucleic acid, e.g., which can confer a desired activity or which can be recombined to produce a desired activity, are recombined to produce a library of recombinant nucleic acids. The library is then screened to identify at least one recombinant nucleic acid that is optimized for the particular property or properties of interest.

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Often, improvements are achieved after one round of recombination and selection. However, recursive sequence recombination can be employed to achieve still further improvements in a desired property, or to bring about new (or "distinct") properties. Recursive sequence recombination entails successive cycles of recombination to generate molecular diversity. That is, one creates a family of nucleic acid molecules showing some sequence identity to each other but differing due to the presence of mutations. In any given cycle, recombination can occur *in vivo* or *in vitro*, intracellularly or extracellularly. Furthermore, diversity resulting from recombination can be augmented in any cycle by applying known methods of mutagenesis (e.g., error-prone PCR or cassette mutagenesis) to either the substrates or products for recombination. In general, however, a single cycle of DNA shuffling of codon-altered nucleic acids provides for generation of surprisingly effective nucleic acids. Accordingly, while recursive approaches to shuffling can be used, single cycle recombination is also preferred. Typically, 2, 3, 4, 5, or even 10 or more cycles of recombination can be performed, each cycle optionally comprising one or more selection steps.

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A recombination cycle is usually followed by at least one cycle of screening or selection for molecules having a desired property or characteristic. If a recombination cycle is performed *in vitro*, the products of recombination, i.e., recombinant segments, are sometimes introduced into cells before the screening step. Recombinant segments can also be linked to an appropriate vector or other regulatory sequences before screening.

55

5 Alternatively, products of recombination generated *in vitro* are sometimes packaged in  
viruses (e.g., bacteriophage) before screening. If recombination is performed *in vivo*,  
10 recombination products can sometimes be screened in the cells in which recombination  
occurred. In other applications, recombinant segments are extracted from the cells, and  
5 optionally packaged as viruses, before screening.

The nature of screening or selection depends on what property or  
15 characteristic is to be acquired or the property or characteristic for which improvement is  
sought, and many examples are discussed below. It is not usually necessary to understand the  
molecular basis by which particular products of recombination (recombinant segments) have  
10 acquired new or improved properties or characteristics relative to the starting substrates. For  
example, a gene can have many component sequences, each having a different intended role  
20 (e.g., coding sequences, regulatory sequences, targeting sequences, stability-conferring  
sequences, subunit sequences and sequences affecting integration). Each of these component  
sequences can be varied and recombined simultaneously. Screening/selection can then be  
25 performed, for example, for recombinant segments that have increased ability to confer  
activity upon a cell without the need to attribute such improvement to any of the individual  
component sequences of the vector.

30 Depending on the particular screening protocol used for a desired property,  
initial round(s) of screening can sometimes be performed using bacterial cells due to high  
20 transfection efficiencies and ease of culture. However, bacterial expression is often not  
practical or desired, and yeast, fungal or other eukaryotic systems are also used for library  
35 expression and screening. Similarly, other types of screening which are not amenable to  
screening in bacterial or simple eukaryotic library cells, are performed in cells selected for  
use in an environment close to that of their intended use. Final rounds of screening can be  
40 performed in the precise cell type of intended use.

If further improvement in a property is desired, at least one and usually a  
45 collection of recombinant segments surviving a first round of screening/selection are subject  
to a further round of recombination. These recombinant segments can be recombined with  
each other or with exogenous segments representing the original substrates or further variants  
30 thereof. Again, recombination can proceed *in vitro* or *in vivo*. If the previous screening step  
identifies desired recombinant segments as components of cells, the components can be

5 subjected to further recombination *in vivo*, or can be subjected to further recombination *in vitro*, or can be isolated before performing a round of *in vitro* recombination. Conversely, if  
10 the previous screening step identifies desired recombinant segments in naked form or as components of viruses, these segments can be introduced into cells to perform a round of *in vivo* recombination. The second round of recombination, irrespective how performed,  
5 generates further recombinant segments which encompass additional diversity that is present in recombinant segments resulting from a previous round (or from multiple previous rounds, e.g., where the process is iteratively repeated).

The second round of recombination can be followed by a further round of  
10 screening/selection according to the principles discussed above for the first round. The stringency of screening/selection can be increased between rounds. Also, the nature of the screen and the property being screened for can vary between rounds if improvement in more  
20 than one property is desired or if acquiring more than one new property is desired. Additional rounds of recombination and screening can then be performed until the  
25 recombinant segments have sufficiently evolved to acquire the desired new or improved property or function.

The practice of this invention involves the construction of recombinant nucleic  
30 acids and the expression of genes in transfected host cells. Molecular cloning techniques to achieve these ends are known in the art. A wide variety of cloning and *in vitro* amplification  
20 methods suitable for the construction of recombinant nucleic acids such as expression vectors are well-known to persons of skill. General texts which describe molecular biological  
35 techniques useful herein, including mutagenesis, include Berger and Kimmel, Guide to Molecular Cloning Techniques, Methods in Enzymology volume 152 Academic Press, Inc., San Diego, CA (Berger); Sambrook et al., Molecular Cloning - A Laboratory Manual (2nd  
40 Ed.), Vol. 1-3, Cold Spring Harbor Laboratory, Cold Spring Harbor, New York, 1989 ("Sambrook") and Current Protocols in Molecular Biology, F.M. Ausubel et al., eds., Current  
25 Protocols, a joint venture between Greene Publishing Associates, Inc. and John Wiley & Sons, Inc., (supplemented through 1998) ("Ausubel"). Methods of transducing cells,  
45 including plant and animal cells, with nucleic acids are generally available, as are methods of expressing proteins encoded by such nucleic acids. In addition to Berger, Ausubel and  
30 Sambrook, useful general references for culture of animal cells include Freshney (Culture of

Animal Cells, a Manual of Basic Technique, third edition Wiley- Liss, New York (1994)) and the references cited therein, Humason (Animal Tissue Techniques, fourth edition W.H. Freeman and Company (1979)) and Ricciardelli, et al., In Vitro Cell Dev. Biol. 25:1016-1024 (1989). References for plant cell cloning, culture and regeneration include Payne et al. (1992) Plant Cell and Tissue Culture in Liquid Systems John Wiley & Sons, Inc. New York, NY (Payne); and Gamborg and Phillips (eds) (1995) Plant Cell, Tissue and Organ Culture: Fundamental Methods Springer Lab Manual, Springer-Verlag (Berlin Heidelberg New York) (Gamborg). A variety of Cell culture media are described in Atlas and Parks (eds) The Handbook of Microbiological Media (1993) CRC Press, Boca Raton, FL (Atlas). Additional information for plant cell culture is found in available commercial literature such as the Life Science Research Cell Culture Catalogue (1998) from Sigma- Aldrich, Inc (St Louis, MO) (Sigma-LSRCCC) and, e.g., the Plant Culture Catalogue and supplement (1997) also from Sigma-Aldrich, Inc (St Louis, MO) (Sigma-PCCS).

Examples of techniques sufficient to direct persons of skill through *in vitro* amplification methods, including the polymerase chain reaction (PCR), the ligase chain reaction (LCR), Q $\beta$ -replicase amplification and other RNA polymerase mediated techniques (e.g., NASBA) are found in Berger, Sambrook, and Ausubel, *id.*, as well as in Mullis *et al.*, (1987) U.S. Patent No. 4,683,202; PCR Protocols A Guide to Methods and Applications (Innis *et al.* eds) Academic Press Inc. San Diego, CA (1990) (Innis); Arnheim & Levinson (October 1, 1990) C&EN 36-47; The Journal Of NIH Research (1991) 3, 81-94; Kwok *et al.* (1989) Proc. Natl. Acad. Sci. USA 86, 1173; Guatelli *et al.* (1990) Proc. Natl. Acad. Sci. USA 87, 1874; Lomell *et al.* (1989) J. Clin. Chem 35, 1826; Landegren *et al.*, (1988) Science 241, 1077-1080; Van Brunt (1990) Biotechnology 8, 291-294; Wu and Wallace, (1989) Gene 4, 560; Barringer *et al.* (1990) Gene 89, 117, and Sooknanan and Malek (1995) Biotechnology 13: 563-564. Improved methods of cloning *in vitro* amplified nucleic acids are described in Wallace *et al.*, U.S. Pat. No. 5,426,039. Improved methods of amplifying large nucleic acids by PCR are summarized in Cheng *et al.* (1994) Nature 369: 684-685 and the references therein, in which PCR amplicons of up to 40kb are generated. One of skill will appreciate that essentially any RNA can be converted into a double stranded DNA suitable for restriction digestion, PCR expansion and sequencing using reverse transcriptase and a polymerase. See, Ausbel, Sambrook and Berger, *all supra*.



5 Oligonucleotides *e.g.*, for use in *in vitro* amplification/ gene reconstruction  
methods, for use as gene probes, or as shuffling targets (*e.g.*, synthetic genes or gene  
10 segments) are typically synthesized chemically according to the solid phase phosphoramidite  
triesther method, *e.g.*, as described by Beaucage and Caruthers (1981), *Tetrahedron Letts.*,  
5 22(20):1859-1862, *e.g.*, using an automated synthesizer, *e.g.*, as described in Needham-  
VanDevanter *et al.* (1984) *Nucleic Acids Res.*, 12:6159-6168 or as is now practiced routinely  
15 in the art. Oligonucleotides can also be custom made and ordered from a variety of  
commercial sources known to persons of skill. Purification of oligonucleotides (*e.g.*, using  
gel-purification methods) to improve the quality of synthesized oligonucleotides can be  
10 particularly desirable in the processes herein to improve the quality of nucleic acid synthesis  
20 protocols.

As noted, essentially any nucleic acid can be custom ordered from any of a  
variety of commercial sources, such as The Midland Certified Reagent Company  
25 (mcrc@oligos.com), The Great American Gene Company (<http://www.genco.com>),  
15 ExpressGen Inc. ([www.expressgen.com](http://www.expressgen.com)), Operon Technologies Inc. (Alameda, CA) and  
many others. Similarly, peptides and antibodies can be custom ordered from any of a variety  
of sources, such as PeptideGenic (pkim@ccnet.com), HTI Bio-products, inc.  
30 (<http://www.htibio.com>), BMA Biomedicals Ltd (U.K.), BioSynthesis, Inc., and many  
others.

#### 20 CODON AND AMINO ACID ALTERED LIBRARIES

35 In the methods of the invention, libraries of codon altered nucleic acids can be  
made and recombined. The codon altered nucleic acids can also include differences in  
encoded amino acid sequences, which can be either conservative or non-conservative in  
40 nature. The codon altered nucleic acids can be derived from a single parental amino acid  
25 sequence, or can be derived from a family of original sequences, *e.g.*, natural or synthetic  
homologous variants of a given sequence. Libraries can exist, *e.g.*, in pools or aliquots of  
cells, viral plaques, enzymatically synthesized pools or aliquots of nucleic acids, or  
45 chemically synthesized pools of nucleic acids. Methods of making libraries of nucleic acids  
are available and taught, *e.g.*, in Berger, Sambrook and Ausubel, *supra*. In one embodiment,  
30 a library as used in the invention comprises at least 2 nucleic acid sequences. In additional

embodiments, the libraries of this invention comprise at least 2, 5, 10, 100, 1000, or more nucleic acid sequences.

As applied to the invention, libraries are typically constructed with a high percentage of codons altered relative to an initial (*e.g.*, wild type) nucleic acid. Codon usage divergence for each of the codon altered nucleic acids can be 50%, 75%, or even 90% or more as compared to the first nucleic acid. This eliminates hybridization to the parental nucleic acid (and thereby inhibits recombination with the parental nucleic acid, a desirable feature in certain embodiments discussed below).

In several embodiments of this invention, codons are modified in members of a gene family so as to increase the degree of identity between the members. In one such embodiment, the genes are homologous genes from different species. In such cases, the degree of nucleic acid identity may be lower than the degree of amino acid identity, at least in part, because of differences in codon usage between the species. In additional embodiments, the homologous genes represent different members of a gene family within a single species. Such genes may encode functionally distinct members of a gene family that nevertheless share significant structural or functional similarity. In preferred embodiments, homologous genes are reverse translated into nucleic acid sequences, and the nucleic acid sequences are modified so as to increase the level of identity between them. Nucleic acids with the modified sequences can then be synthesized *in vitro*. In particularly preferred embodiments, the modified nucleic acid sequences are at least as identical to each other as the original amino acid sequences.

Additional sequence diversity is provided by generating nucleic acids with non-overlapping non-conservative substitutions in each of the codon altered nucleic acids as compared to the first nucleic acid. This provides for reversion to wild-type upon recombination, while optionally allowing for the incorporation of non-conservative changes to the sequence in the event that they produce a detectable improvement during screening.

Modification of the codons of one or more of the codon altered nucleic acids to provide one or more different hydrophobic core residue for an encoded polypeptide as compared to the first polypeptide is also provided. This modification of core amino acids provides minor differences in encoded proteins, while changing the mutational spectrum of the resulting nucleic acid, thereby increasing sequence diversity.

5 In addition, due to the constraints of the translational machinery for a given  
cell, codon usage may need to be altered when expressed sequences are shuttled between  
different organisms (e.g., animal cells, plant cells, bacterial cells, etc.) for optimal expression.  
10 This produces a nucleic acid which encodes the same protein, but which, after typical forms  
of point mutation, will access a different mutational diversity than the original form of the  
protein.

15 In one embodiment, phage libraries are made and recombined in mutator  
strains such as cells with mutant or impaired gene products of *mutS*, *mutT*, *mutH*, *mutL*,  
*ovrD*, *dcm*, *vsr*, *umuC*, *umuD*, *sbcB*, *recJ*, etc. The impairment is achieved by genetic  
20 mutation, allelic replacement, selective inhibition by an added reagent such as a small  
compound or an expressed antisense RNA, or other techniques. High multiplicity of  
infection (MOI) libraries are used to infect the cells to increase recombination frequency.  
Additional strategies for making phage libraries and or for recombining DNA from donor and  
25 recipient cells are set forth in U.S. Pat. No. 5,521,077. Additional recombination strategies  
for recombining plasmids in yeast are set forth in WO 97 07205.

30 The library to be made can be an *in vitro* set of molecules, or present in cells,  
phage or the like. Virtual libraries of nucleic acids generated in silico are also a feature of the  
invention (see also, Selifonov and Stemmer, *supra*). Generally, the library is screened to  
identify at least one recombinant nucleic acid that exhibits distinct or improved activity  
20 compared to the parental nucleic acid or nucleic acids which are recombined. Additional  
details on making appropriate libraries are found below, e.g., in the section entitled "Formats  
for Sequence Recombination."

#### TARGETS FOR CODON MODIFICATION AND SHUFFLING

40 Essentially any nucleic acid can be codon altered and shuffled. No attempt is  
made herein to identify the hundreds of thousands of known nucleic acids. Common  
sequence repositories for known proteins include GenBank EMBL, DDBJ and the NCBI.  
Other repositories can easily be identified by searching the internet.

45 One class of preferred targets for activation includes nucleic acids encoding  
therapeutic proteins such as erythropoietin (EPO), insulin, peptide hormones such as human  
30 growth hormone; growth factors and cytokines such as epithelial Neutrophil Activating  
Peptide-78, GRO $\alpha$ /MGSA, GRO $\beta$ , GRO $\gamma$ , MIP-1 $\alpha$ , MIP-1 $\beta$ , MCP-1, epidermal growth  
50

factor, fibroblast growth factor, hepatocyte growth factor, insulin-like growth factor, the interferons, the interleukins, keratinocyte growth factor, leukemia inhibitory factor, oncostatin M, PD-ECSF, PDGF, pleiotropin, SCF, c-kit ligand, VEGF, G-CSF etc. Many of these proteins are commercially available (See, e.g., the Sigma BioSciences 1997 catalogue and price list), and the corresponding genes are well-known.

Another class of preferred targets are transcriptional and expression activators.

Example transcriptional and expression activators include genes and proteins that modulate cell growth, differentiation, regulation, or the like. Expression and transcriptional activators are found in prokaryotes, viruses, and eukaryotes, including fungi, plants, and animals, including mammals, providing a wide range of therapeutic targets. It will be appreciated that expression and transcriptional activators regulate transcription by many mechanisms, e.g., by binding to receptors, stimulating a signal transduction cascade, regulating expression of transcription factors, binding to promoters and enhancers, binding to proteins that bind to promoters and enhancers, unwinding DNA, splicing pre-mRNA, polyadenylating RNA, and degrading RNA. Expression activators include cytokines, inflammatory molecules, growth factors, their receptors, and oncogene products, e.g., interleukins (e.g., IL-1, IL-2, IL-8, etc.), interferons, FGF, IGF-I, IGF-II, FGF, PDGF, TNF, TGF- $\alpha$ , TGF- $\beta$ , EGF, KGF, SCF/c-Kit, CD40L/CD40, VLA-4/VCAM-1, ICAM-1/LFA-1, and hyalurin/CD44; signal transduction molecules and corresponding oncogene products, e.g., Mos, Ras, Raf, and Met; and transcriptional activators and suppressors, e.g., p53, Tat, Fos, Myc, Jun, Myb, Rel, and steroid hormone receptors such as those for estrogen, progesterone, testosterone, aldosterone, the LDL receptor ligand and corticosterone.

Similarly, proteins from infectious organisms for possible vaccine applications, described in more detail below, including infectious fungi, e.g., *Aspergillus*, *Candida* species; bacteria, particularly *E. coli*, which serves a model for pathogenic bacteria, as well as medically important bacteria such as *Staphylococci* (e.g., *aureus*), *Streptococci* (e.g., *pneumoniae*), *Clostridia* (e.g., *perfringens*), *Neisseria* (e.g., *gonorrhoea*), *Enterobacteriaceae* (e.g., *coli*), *Helicobacter* (e.g., *pylori*), *Vibrio* (e.g., *cholerae*), *Campylobacter* (e.g., *jejuni*), *Pseudomonas* (e.g., *aeruginosa*), *Haemophilus* (e.g., *influenzae*), *Bordetella* (e.g., *pertussis*), *Mycoplasma* (e.g., *pneumoniae*), *Ureaplasma* (e.g., *urealyticum*), *Legionella* (e.g., *pneumophila*), *Spirochetes* (e.g., *Treponema*, *Leptospira*, and *Borrelia*),

5 *Mycobacteria* (e.g., *tuberculosis*, *smegmatis*), *Actinomyces* (e.g., *israelii*), *Nocardia* (e.g.,  
*asteroides*), *Chlamydia* (e.g., *trachomatis*), *Rickettsia*, *Coxiella*, *Ehrlichia*, *Rochalimaea*,  
10 *Brucella*, *Yersinia*, *Francisella*, and *Pasteurella*; protozoa such as sporozoa (e.g., *Plasmodia*),  
*rhizopods* (e.g., *Entamoeba*) and flagellates (*Trypanosoma*, *Leishmania*, *Trichomonas*,  
5 *Giardia*, etc.); viruses such as ( + ) RNA viruses (examples include Poxviruses e.g., *vaccinia*;  
Picornaviruses, e.g. *polio*; Togaviruses, e.g., *rubella*; Flaviviruses, e.g., HCV; and  
15 Coronaviruses), ( - ) RNA viruses (examples include Rhabdoviruses, e.g., VSV;  
Paramyxoviruses, e.g., RSV; Orthomyxoviruses, e.g., influenza; Bunyaviruses; and  
Arnaviruses), dsDNA viruses (Reoviruses, for example), RNA to DNA viruses, i.e.,  
10 Retroviruses, e.g., especially HIV and HTLV, and certain DNA to RNA viruses such as  
20 Hepatitis B virus.

Other proteins relevant to non-medical uses, such as inhibitors of transcription  
or toxins of crop pests e.g., insects, fungi, weed plants, and the like, are also preferred targets  
25 for shuffling. Industrially important enzymes such as monooxygenases, proteases, nucleases,  
15 and lipases are also preferred targets. As an example, subtilisin can be evolved by shuffling  
codon altered forms of the gene for subtilisin (von der Osten et al., *J. Biotechnol.* 28:55-68  
(1993) provide a subtilisin coding nucleic acid). Proteins which aid in folding such as the  
30 chaperonins are also preferred.

Preferred known genes suitable for codon alteration and shuffling also include  
20 the following: Alpha-1 antitrypsin, Angiostatin, Antihemolytic factor, Apolipoprotein,  
35 Apoprotein, Atrial natriuretic factor, Atrial natriuretic polypeptide, Atrial peptides, C-X-C  
chemokines (e.g., T39765, NAP-2, ENA-78, Gro-a, Gro-b, Gro-c, IP-10, GCP-2, NAP-4,  
SDF-1, PF4, MIG), Calcitonin, CC chemokines (e.g., Monocyte chemoattractant protein-1,  
Monocyte chemoattractant protein-2, Monocyte chemoattractant protein-3, Monocyte  
40 inflammatory protein-1 alpha, Monocyte inflammatory protein-1 beta, RANTES, I309,  
25 R83915, R91733, HCC1, T58847, D31065, T64262), CD40 ligand, Collagen, Colony  
stimulating factor (CSF), Complement factor 5a, Complement inhibitor, Complement  
receptor 1, Factor IX, Factor VII, Factor VIII, Factor X, Fibrinogen, Fibronectin,  
45 Glucocerebrosidase, Gonadotropin, Hedgehog proteins (e.g., Sonic, Indian, Desert),  
30 Hemoglobin (for blood substitute; for radiosensitization), Hirudin, Human serum albumin,  
Lactoferrin, Luciferase, Neurturin, Neutrophil inhibitory factor (NIF), Osteogenic protein,  
50

5 Parathyroid hormone, Protein A, Protein G, Relaxin, Renin, Salmon calcitonin, Salmon  
growth hormone, Soluble complement receptor I, Soluble I-CAM 1, Soluble interleukin  
10 receptors (IL-1, 2, 3, 4, 5, 6, 7, 9, 10, 11, 12, 13, 14, 15), Soluble TNF receptor,  
Somatomedin, Somatostatin, Somatotropin, Streptokinase, Superantigens, i.e.,  
5 Staphylococcal enterotoxins (SEA, SEB, SEC1, SEC2, SEC3, SED, SEE), Toxic shock  
syndrome toxin (TSST-1), Exfoliating toxins A and B, Pyrogenic exotoxins A, B, and C, and  
15 M. arthritides mitogen, Superoxide dismutase, Thymosin alpha 1, Tissue plasminogen  
activator, Tumor necrosis factor beta (TNF beta), Tumor necrosis factor receptor (TNFR),  
Tumor necrosis factor-alpha (TNF alpha) and Urokinase. Many other known coding nucleic  
20 acids, such as those in Genebank™, can be codon-altered and shuffled.

GENES WITH CODON USAGE REDESIGNED AND CHEMICALLY SYNTHESIZED  
AS STARTING MATERIALS FOR GENE FAMILY SHUFFLING--EXPANDING THE  
DIVERSITY OF DNA SHUFFLING.

25 Because the genetic coding preference among organisms ranges from quite  
15 similar to very different, homologous genes from different organisms can have significantly  
lower homology at the nucleic acid level than at the amino acid level. For example, genetic  
information for some bacterial species is high in GC content (up to 70%), while others have  
30 AT rich (>60%) codon usage. Thus, genes from different organisms may have, for example,  
40-60% amino acid identity but only 25-35% nucleic acid identity. It is often desirable to  
20 increase such levels of nucleic acid identity so as to enhance the ability of the homologous  
sequences to recombine, thereby increasing the efficiency of family shuffling using the  
35 methods of this invention. In other aspects, it is actually preferable to decrease the rate of  
recombination in a system, e.g., when using vectors it is sometimes desirable to decrease the  
rate of recombination between the vector and the host DNA, thereby increasing the safety of  
40 the vector. The following examples address specific issues with regard to shuffling codon  
25 altered nucleic acids.

Altering codon usage to increase homology

45 In one aspect, protein sequences of gene family members are reverse  
translated back into DNA sequences, for example by using one of the preferable codon usage  
30 charts in any conventional DNA manipulation program (e.g. the Wisconsin Package™,  
SeqWeb, OMIGA, SeqApp, SeqPup, MacVector, DNA stryder, GeneWorks, etc.). The  
50

choice of codon usage is often determined by the host in which the genes will be expressed. After maximizing the percentage of DNA sequence identity, the genes are chemically synthesized, *e.g.*, using a high throughput oligonucleotide synthesizer in, *e.g.* a 96-well format, optionally in conjunction with polymerase and/ or ligase gene synthesis methods.

In general, the DNA sequence similarity after such treatment will be at least as high as the amino acid similarity, but can be at least about 10% to 15% *higher* than the amino acid identity (in contrast to the situation for naturally occurring genes, which are ordinarily less well conserved than encoded polypeptides), based on the random frequency of sequence identity for any given codon. In most cases, the minimal requirement for amino acid identity can be as low as about 35% while still retaining adequate nucleic acid homology for standard recombination methods (as discussed, *supra*, oligonucleotide-mediated recombination methods do not require high levels of similarity to achieve recombination). In some cases, however, the minimal amino acid identity can be even lower, *e.g.* if the conserved regions are clustered within the genes.

Example: Shuffling codon-modified EPO

The protein erythropoietin alpha, also known as EPO, Epogen, and Procrit is a hematopoietic hormone, providing a variety of benefits to patients suffering from anemia (a common symptom of, *e.g.*, AIDS). EPO is produced as a pharmaceutical, with sales of nearly 1 billion dollars world-wide. Accordingly, proteins with EPO-like activity (and preferably superior activity) are of substantial commercial interest.

Figure 1 shows the sequence of a part of the monkey EPO gene, which is similar to the human EPO gene. Figure 2 shows an example of a codon altered EPO nucleic acid (or "wobble" EPO gene). In general, transversions rather than transition mutations are made where possible. The purpose of this strategy is to maximally disrupt hybridization of the resulting gene with naturally occurring EPOs. Figure 3 shows an alignment of naturally occurring EPOs.

This strategy is further fine-tuned by applying standard rules of base pairing (*e.g.*, elimination of G-C pairing and GC stacking) to maximize sequence disruption; in addition, conservative or non-conservative amino acid modifications can also be made (in some cases, where multiple codon-altered nucleic acids are shuffled, it is desirable to make codon altered nucleic acids with non-overlapping non-conservative substitutions to permit

5                   reversion to the wild-type amino acid during shuffling). The size of the sequence space for  
nucleic acids encoding EPO is large, at about  $2.8 \times 10^{88}$  different sequences (there are about  
10                    $10^{80}$  particles in the universe; thus, it is physically impossible to make all of the possible  
sequences encoding EPO). As indicated schematically in Figure 4, if one only considers the  
5                   maximally divergent wobble genes (those that use the alternative types of codons for leucine,  
arginine, and serine), there is still a sequence space of  $10^{38}$  sequences encoding EPO. The  
15                   overall strategy is to synthesize a library of wobble genes, screen for expression and activity  
and DNA shuffle desirable genes as desired (e.g., by recursive processes).

                  It is of interest to further evolve codon-altered nucleic acids. Shuffling with  
10                   other homologous genes from nature, designed genes (incorporating libraries of designed  
sequence variation), and genes containing mutations of interest are strategies for evolving  
any gene of interest. However, the codon altered nucleic acid may not be easily shuffled with  
these genes because of the sequence differences; or they may be undesirable for other reasons  
25                   (e.g., the naturally occurring sequences may be proprietary, or include proprietary elements).  
15                   These difficulties can be avoided by synthesizing codon-altered homologous nucleic acids  
which encode desired amino acid variations (e.g., those found in homologous genes), but  
which have a codon-set close to the nucleic acid(s) to be recombined (thereby permitting,  
30                   e.g., hybridization during recombination).

                  For example, after identifying homologues of interest (e.g., those shown in  
20                   Figure 3 for EPO), codon-altered nucleic acids encoding the same proteins are synthesized  
with a similar codon selection. Standard family shuffling is then practiced with the codon  
35                   altered nucleic acids. This is shown schematically for EPO in Figure 5.

                  EPO wobble variants are screened for expression and then receptor binding  
assays are conducted in an ELISA format, using human EPOr-Fc fusions. Following  
40                   25                   selection of binding variants, activity is measured as thymidine incorporation in UT7-EPO (A  
human bone marrow cell line) cell proliferation assays. Cells are treated for 2-3 days with  
various concentration of EPO variants after which time they are incubated in the presence of  
45                   3-H thymidine for 4 hours and incorporation of thymidine is measured. *See also*, Erickson-  
miller et al. (1997) Blood 90:2421 (for the receptor binding assay), and Wen et al. (1994) J.  
30                   Biol. Chem. 269:22839-22846 (for the thymidine incorporation assay).



Assays for selecting EPO can also be based, e.g., on the ability of EPO proteins to stimulate the growth of blood cell, e.g., in vitro or in vivo.

Example: Codon Shuffling G-CSF

Family shuffling can be used to breed diversity from genes into the libraries to be screened. Additionally, design heuristics such as randomization of hydrophobic core residues can be used to take advantage of the redundancy between primary structure and tertiary structure of proteins (i.e. many different primary structures encode proteins with very similar three dimensional structures).

Design heuristics are employed to create a sequence space of mutants that are predicted to be highly biased (relative to random mutagenesis) to encode proteins which preserve the original activity. Methods such as high throughput (HTP) screening and phage panning are used to identify members of the designed libraries that have the desired activity. DNA shuffling is used to breed this population of active clones in order to fine tune the mutants, thus allowing one to evolve variants with equivalent or superior function relative to the naturally occurring proteins.

Figures 6 and 7 show several mammalian homologues of G-CSF. Figure 8 shows the hydrophobic core residues of human G-CSF (blackened out). Figure 9 shows a strategy for evolving variants of human G-CSF that are highly divergent in sequence. First, three genes are synthesized (Genes 1, 2 and 3, Figure 8) which contain all of the mammalian homologue diversity of G-CSF. These genes are shuffled, phage panned against the G-CSF receptor, and HTP screened for biological function (receptor activation). Active clones are iteratively shuffled and screened if necessary to give evolved variants that rival or surpass the human gene in activity (on human cells).

Next, one evolves a variant that has a highly mutated hydrophobic core. This is schematically illustrated in Figure 8, and the specific strategy for performing the biological screening is schematically illustrated in Figure 9. It is expected that the best mutants obtained after screening hydrophobic core randomized libraries may be less active than wild type human G-CSF because it is difficult to initially optimize activity in such a procedure. Family shuffling is used to obtain optimized variants. This is done by synthesizing genes which contain mammalian homologue diversity at all but the hydrophobic core positions; but

5 they are synthesized in the context of an evolved, non-wild type hydrophobic core. Family shuffling is used to optimize around the new hydrophobic core.

10 This strategy works because there are functionally similar hydrophobic cores for wild type proteins that consist of largely different amino acids than the wild type protein.

5 This understanding is supported by recent experiments in model systems. For example, 53% of randomized sequences for three residues in the hydrophobic core of lambda repressor are folded and biologically active (Lim and Sauer (1991) J. Mol. Biol. 219:359-376). Protein design by patterning of polar and non-polar amino acids, where 24 residues in the hydrophobic core of a 4-helix bundle protein were randomized by Kamtekar et al. (1993) Science 262:1321. Folded, alpha helical proteins were recovered from about 1% of the clones. Desjarlais and Handel (1995) Current Opinion in Biotechnology 6:460-466 showed a mutant of Rop, another 4-helix bundle protein, where four hydrophobic core residues have been randomized and active mutants have been obtained. Axe et al. (1996) PNAS 95:5590-5594 showed that randomizing 13 hydrophobic core residues in the enzyme barnase resulted in 23% of the clones in the library retaining biological activity. Gassener et al. (1996) PNAS 93:12155-12158 describe a mutant of T4 lysozyme where 10 residues in the hydrophobic core are replaced with Met. This is taken as evidence that the hydrophobic core of this protein is very tolerant to substitution. Taken together, this experimental evidence on model systems shows that the hydrophobic cores of many proteins can be replaced with other hydrophobic residues that pack in a similar fashion to give an active protein. This degeneracy is exploited to evolve novel forms of natural and codon-altered genes.

35 A related approach is to search the protein databases for a protein that has a similar activity to a protein that one wishes to evolve. Denesyuk et al. (1996) J. Theor. Biol. shows the results of such a search for G-CSF. LIF is a very similarly folded protein. One can use LIF as a 'scaffold' on which to place residues of G-CSF that are required for activity. Given LIF with a G-CSF "toupee," one would family shuffle the LIF scaffold so as to obtain a variant in which the toupee is displayed in a fully biologically active form.

45 Another approach is to use computational methods to create families of variants that are predicted to be functional. Dahiyat and Mayo Science recently described computer methods that are used to design proteins. Proteins are simulated on the computer, often with the aid of genetic algorithms, and a subset that are deemed 'fit' are actually

5 synthesized and 'analyzed'. These computational methods are becoming increasingly  
powerful. They would be useful to, for example, predict a family of mutations on the surface  
10 of a protein that would not destroy function. DNA shuffling can be used to optimized active  
clones obtained by design. Taking the example of G-CSF, one could use computational  
5 methods in combination with all structure function data (for example alanine scan data for G-  
CSF reported recently by Reidhaar-Olsen in Biochemistry) to design a family of putatively  
15 functional variants. One could, for example, design the family to have minimal DNA identity  
to the wild type gene given the design constraints. This library is synthesized, put through  
biological screens and/or selections (i.e. panning against the G-CSF receptor), and active  
10 variants are obtained. DNA shuffling is then used to evolve these active variants to have the  
desired level of function.

G-CSF proteins are displayed on phage and screened for binding to human G-  
CSF receptor in an ELISA format. Variants that bind receptor are selected in a high  
25 throughput screen for receptor activation. This cell based assay measures receptor activation  
via a reporter gene (such as luciferase) activated by a G-CSF responsive construct containing  
15 STAT binding elements. Cells (such as HepG2) are transformed with a G-CSF responsive  
reporter plasmid and treated with the codon shuffled G-CSF variant for 2.5 hours. Cells are  
30 then lysed and luciferase activity measured. *See also*, Tian et al (1998) Science 281:257-259.

Example: Codon Shuffling Alkaline Phosphatase

20 Alkaline phosphatase is a widely used reporter enzyme for ELISA assays,  
protein fusion assays, and in a secreted form as a reporter gene for mammalian cells. A more  
35 active form of the enzyme is desirable.

A codon altered form of alkaline phosphatase was generated by PCR assembly  
using the oligos set forth in Figure 10. A map of the oligos is set forth in Figure 11. The  
40 25 procedure used was essentially identical to that taught in Stemmer et al. (1994) Gene 164:49-  
57. In brief, the oligos were mixed 1:1 at a variety of dilutions and PCR assembled by  
performing e.g., 25-60 cycles of PCR at e.g., 94 °C (60 sec.), 94 °C (30 sec.), 50 °C (30 sec.),  
45 72 °C (30 sec). Assembly of the BIAP gene was conducted in a circular format and gene  
fragments were purified. ~100,000 colonies were screened on LB/amp plates (~1/10 are wt  
30 plasmid). About 1/10 showed a bluer color than background. Plasmid DNA showed a  
correct insertion.

5 In general, petri-dish screening using the typical colorimetric assay for  
phosphatase activity can be used for screening. This has the advantage of being simple, high  
throughput, and semi quantitative. Microtiter plate screening, also preferred, is colorimetric,  
10 and quantitative, although additional instrumentation can be required for implementation.

5 Example: Codon Shuffling to Reduce Competent Virus Production from  
Vectors and to Generate Attenuated Viruses as Immunogenic Compositions  
and Vaccines

15 Cells can be stably transduced with a number of viral vectors including those  
derived from retroviruses, pox viruses, adenoviruses (Ads), herpes viruses and parvoviruses.

10 Common viral vectors include those derived from murine leukemia viruses (MuLV), gibbon  
ape leukemia viruses (GaLV), human immuno deficiency viruses (HIV), adenoviruses, adeno  
20 associated viruses (AAVs), Epstein Barr viruses, canarypox viruses, cowpox viruses, and  
vaccinia viruses. Viral vectors based upon retroviruses, adeno-associated viruses, herpes  
viruses and adenoviruses are all used as gene therapy vectors for the introduction of  
25 therapeutic nucleic acids into the cells of an organism by *ex vivo* and *in vivo* methods.

When using viral vectors, packaging cells are commonly used to prepare  
virions used to transduce target cells. In these vectors, trans-active genes are rendered  
30 inactive and "rescued" by trans-complementation to provide a packaged vector. This form of  
trans complementation is provided by co-infection of a packaging cell with a virus or vector  
20 which supplies functions missing from a particular gene therapy vector in trans, or by using a  
cell line (e.g., 293 cells) which have viral components integrated into the genome of the  
35 packaging cell. For instance, cells transduced with HIV or murine retroviral proviral  
sequences which lack the nucleic acid packaging site produce retroviral trans active  
components, but do not specifically incorporate the retroviral nucleic acids into the capsids  
25 produced, and therefore produce little or no live virus.

If these transduced "packaging" cells are subsequently transduced with a  
vector nucleic acid which lacks coding sequences for retroviral trans active functions, but  
includes a packaging signal, the vector nucleic acid is packaged into an infective virion. A  
45 number of packaging cell lines useful for MoMLV-based vectors are known in the art, such  
30 as PA317 (ATCC CRL 9078) which expresses MoMLV core and envelope proteins see,  
Miller et al. *J. Virol.* 65:2220-2224 (1991). Carrol et al. (1994) *Journal of virology*  
68(9):6047-6051 describe the construction of packaging cell lines for HIV viruses.

Reciprocal complementation of defective HIV molecular clones is described, e.g., in Lori et al. (1992) Journal of Virology 66(9) 5553-5560.

Functions of viral replication not supplied by trans-complementation which are necessary for replication of the vector are present in the vector. In HIV, this typically includes, e.g., the TAR sequence, the sequences necessary for HIV packaging, the RRE sequence if the instability elements of the p17 gene of gag is included, and sequences encoding the polypurine tract. HIV sequences that contain these functions include a portion of the 5' long terminal repeat (LTR) and sequences downstream of the 5' LTR responsible for efficient packaging, i.e., through the major splice donor site ("MSD"), and the polypurine tract upstream of the 3' LTR through the U3R section of the 3' LTR. The packaging site (psi site or  $\psi$  site) is partially located adjacent to the 5' LTR, primarily between the MSD site and the gag initiator codon (AUG) in the leader sequence. See, Garzino-Demo et al. (1995) Hum. Gene Ther. 6(2): 177-184. For a general description of the structural elements of the HIV genome, see, Holmes et al. PCT/EP92/02787.

Another common vector is based upon adenovirus. Typically, vectors which include the adenovirus ITRs (Gingeras *et al.* (1982) *J. Biol. Chem.* 257:13475-13491) are packaged in, e.g., 293 cells, which provide many of the components necessary for vector packaging.

Adeno-associated viruses (AAVs) utilize helper viruses such as adenovirus or herpes virus to achieve productive infection. In the absence of helper virus functions, AAV integrates (site-specifically) into a host cell's genome, but the integrated AAV genome has no pathogenic effect. The integration step allows the AAV genome to remain genetically intact until the host is exposed to the appropriate environmental conditions (*e.g.*, a lytic helper virus), whereupon it re-enters the lytic life-cycle. Samulski (1993) *Current Opinion in Genetic and Development* 3:74-80 and the references cited therein provides an overview of the AAV life cycle. For a general review of AAVs and of the adenovirus or herpes helper functions *see*, Berns and Bohensky (1987) *Advanced in Virus Research*, Academic Press., 32:243-306. The genome of AAV is described in Laughlin *et al.* (1983) *Gene*, 23:65-73. Expression of AAV is described in Beaton *et al.* (1989) *J. Virol.*, 63:4450-4454. In general, the packaging sites for all parvoviruses, including B 19 and AAV are located in the viral ITRs. Recombinant AAV vectors (rAAV vectors) deliver foreign nucleic acids to a wide

5 range of mammalian cells (Hermonat & Muzycka (1984) *Proc Natl Acad Sci USA* 81:6466-6470; Tratschin *et al.* (1985) *Mol Cell Biol* 5:3251-3260), integrate into the host chromosome (McLaughlin *et al.* (1988) *J Virol* 62: 1963-1973), and show stable expression of the  
10 transgene in cell and animal models (Flotte *et al.* (1993) *Proc Natl Acad Sci USA* 90:10613-10617). rAAV vectors are able to infect non-dividing cells (Podsakoff *et al.* (1994) *J Virol* 68:5656-66; Flotte *et al.* (1994) *Am. J. Respir. Cell Mol. Biol.* 11:517-521). Further  
15 advantages of rAAV vectors include the lack of an intrinsic strong promoter, thus avoiding possible activation of downstream cellular sequences, and the vector's naked icosohedral capsid structure, which renders the vectors stable and easy to concentrate by common  
20 laboratory techniques.

One problem with previously existing vector packaging strategies is that  
vectors to be packaged can recombine with nucleic acids providing packaging functions in  
trans, producing a replication-competent virus. This can be a problem both when vectors are  
25 produced for therapeutic applications (e.g., in gene therapy) and during production of encoded components in vitro. The present invention provides a way of reducing or  
15 eliminating recombination between nucleic acids encoding trans-active components and vector nucleic acids encoding packaging sites.

In particular, nucleic acid subsequences of a vector which are adjacent to  
modified or deleted elements provided in trans, are codon modified to eliminate hybridization  
20 to wild-type sequences. Because these sequences do not hybridize, they cannot recombine with nucleic acids producing trans-active components. One additional advantage of this  
35 approach is that the vectors also cannot recombine with live viruses, e.g., in a human body which is infected with a virus that packages vector elements. As noted above, two types of gene therapy vectors are those based upon retroviruses (which can be packaged by, e.g., HIV-  
40 25 1) and adenoviruses (which can be packaged by adenovirus).

Alternatively, the nucleic acids encoding trans-active components can be  
codon modified so that they do not hybridize to wild-type sequences. This also prevents  
45 recombination with vectors having wild-type sequences, preventing recombination and formation of replication competent viruses.

5 After codon modification, vectors or trans active nucleic acids can be shuffled as described *supra*, and screened for the ability to package nucleic acids, or to be packaged, as appropriate.

10 It will be appreciated that codon modification of viral sequences has an additional use as well. Codon alteration of viral sequences can result in attenuation of the virus, e.g., due to modification of regulatory sequences, alterations in mRNA secondary structure, inefficient translation due to rare codon use, and the like. Such "codon attenuated" viruses have a significant advantage over existing attenuated viruses (which are typically generated by serial passage in cells other than the normal host type for the virus). In particular, codon attenuated viruses can encode a wild-type set of proteins, making them ideal as immunogenic compositions to generate antibodies, or to use as vaccines. Viral proteins can also be used in various diagnostic assays. For example, the standard diagnostic test for HIV infection in current use tests for the presence of anti-HIV antibodies in blood by probing with viral proteins.

15 Example: Codon Usage Libraries to evolve Functional Variants with reduced Recombination With Natural Gene Sequences—Adenovirus

Adenovirus is a common vector used, e.g., for gene therapy. The virus is typically modified to make it replication deficient. This can be achieved e.g., by deleting the E1 and E4 genes. The functions of E1 and E4 can be supplied by trans complementation when E1 and E4 deleted vectors are grown in the ubiquitous human embryonic kidney cell line 293, which has uncharacterized adenovirus fragments incorporated into their genome that supply the missing functions in trans. The replication defective adenoviral vectors recombine at a low, but clinically significant frequency, resulting in replication competent adenovirus contamination of vector preparations. Because adenovirus has detrimental effects on health, this is a significant problem for application of adenovirus-based gene therapy vectors.

In the present invention, a codon usage library encompassing several hundred bases to several kilobases of sequence flanking the adenovirus E1 and E4 genes are made. The library is designed to enforce a high degree of divergence from the natural adenoviral consensus sequence, while at the same time incorporating a large degree of degeneracy in the codons to allow for a large space of sequence diversity to be searched. The design principle

is to obtain mutants that encode the same or similar protein sequence, but with many mismatches to the wild-type E1 and E4 sequences found in the 293 genome. These mismatches strongly reduce the frequency of unwanted recombination with the trans complementary genes. Consequently, engineered adenoviral vectors, or adenovirus helper vectors which package adenoviral sequences which include packaging sequences (adenoviral or adeno-associated viral ITRs) in trans have reduced levels of recombination. This provides for a lower rate of competent adenovirus production, making culture and production of such vectors safer.

Evolution Impaired Viruses Created by Massive Codon Usage Alteration As a General Approach to Vaccines--HIV

HIV-1 and HIV-2 are genetically related, antigenically cross reactive, and share a common cellular receptor (CD4). See, Rosenberg and Fauci (1993) in *Fundamental Immunology, Third Edition* Paul (ed) Raven Press, Ltd., New York (Rosenberg and Fauci 1) and the references therein for an overview of HIV infection. HIV-1 infection is epidemic world wide, causing a variety of immune system-failure related phenomena commonly termed acquired immune deficiency syndrome (AIDS). HIV type 2 (HIV-2) has been isolated from both healthy individuals and patients with AIDS-like illnesses (Andreasson, *et al.* (1993) *Aids* 7, 989-93; Clavel, *et al.* (1986) *Nature*, 324, 691-695; Gao, *et al.* (1992) *Nature* 358, 495-9; Harrison, *et al.* (1991) *Journal of Acquired Immune Deficiency Syndromes* 4, 1155-60; Kanki, *et al.* (1992) *American Journal of Epidemiology* 136, 895-907; Kanki, *et al.* (1991) *Aids Clinical Review* 1991, 17-38; Romieu, *et al.* (1990) *Journal of Acquired Immune Deficiency Syndromes* 3, 220-30; Naucier, *et al.* (1993) *International Journal of STD and Aids* 4, 217-21; Naucier, *et al.* (1991) *Aids* 5, 301-4). Although HIV-2 AIDS cases have been identified principally from West Africa, sporadic HIV-2 related AIDS cases have also been reported in the United States (O'Brien, *et al.* (1991) *Aids* 5, 85-8) and elsewhere. HIV-2 will likely become endemic in other regions over time, following routes of transmission similar to HIV-1 (Harrison, *et al.* (1991) *Journal of Acquired Immune Deficiency Syndromes* 4, 1155-60; Kanki, *et al.* (1992) *American Journal of Epidemiology* 136, 895-907; Romieu, *et al.* (1990) *Journal of Acquired Immune Deficiency Syndromes* 3, 220-30). Epidemiological studies suggest that HIV-2 produces human disease with lesser penetrance than HIV-1, and exhibits a considerably longer period of clinical latency (at least



25 years, and possibly longer, as opposed to less than a decade for HIV-1; see, Kanki, *et al.* (1991) *Aids Clinical Review* 1991, 17-38; Romieu, *et al.* (1990) *Journal of Acquired Immune Deficiency Syndromes* 3, 220-30, and Travers *et al.* (1995) *Science* 268: 1612-1615).

The ability of HIV virus populations to rapidly point mutate to avoid the immune response poses a special challenge for vaccine design. While the immune system has responded to viruses in a gradual and co-evolutionary manner, the present invention provides a general approach that provides for massively faster evolution to produce new vaccines to stimulate more effective immune responses.

For example, during the incubation period for HIV infection, which lasts for several years, low titers of HIV can result from high HIV replication rates in conjunction with efficient viral clearance by the immune system. In response to these selective forces, virus mutations are selected which reduce recognition and neutralization by the immune system's B and T-cell responses. See, Lukashov *et al.* (1995) *J. Virol.* 69:6911-6916. During the long incubation time, these mutations accumulate and eventually overwhelm the immune system's defenses. See, Ho *et al.* (1995) *Nature*.

Live attenuated vaccines, typically produced by prolonged growth of human viruses in animal cells, have proven useful as vaccines for several diseases, including mumps, rubella and measles. Attenuation involves the slow accumulation of many mutations throughout the viral genome during the course of adapting to growth in the animal cells. When used to vaccinate humans, the attenuated virus grows only weakly and elicits a complex immune response which the virus is unable to avoid. The mutations in the attenuated virus could, in principle, revert in the same stepwise fashion that it underwent to grow in culture.

The risk of reversion is highest in viruses with a high mutation rate such as HIV-1, which makes this strategy dangerous under current techniques for vaccine development. It is worth noting, however, that protective effects against HIV-1 are observed following infection with the related HIV-2 virus, which is much less pathogenic than HIV-1. Thus, protective effects against HIV can be achieved with live vaccines.

To reduce the risk of reversion, a large number of mutations need to accumulate in the virus. However, if too many mutations are present, the immune system in

5 effect recognizes the attenuated virus, but not the virus against which a protective effect is sought.

10 As provided herein, immunogenic compositions such as vaccines are created which contain a large number of silent substitutions. In contrast to existing attenuated  
5 viruses, such viruses have native protein sequences and elicit essentially the same immune responses as the corresponding wild-type virus (typically one or a few additional disabling mutations can also be incorporated). Codon alteration results in two effects that both  
15 increase the potential of the vaccine.

First, like standard attenuated viruses, the growth of codon-altered viruses is  
10 attenuated, due to the effect of the codon alterations on translation, regulatory sequences, mRNA folding, packaging, and the like. For example, regulation of HIV-1 envelope  
20 expression has been observed as a result of codon usage. See, Haas et al. (1989) Current Biology 6(3):315-324.

25 Second, codon alteration results in impairment of virus evolution. As discussed above, modification of the codons alters the mutational escape spectrum of the virus, upsetting the evolutionary selection for specific codons.

30 The six codon amino acids are the best targets for codon alteration. Serine, arginine and leucine each have one group of four codons, plus two codons in an unrelated group. See, Figure 12. Switching all of the serine codons from AGY to TCX and vice versa,  
20 yields proteins with unaltered amino acid sequences. See also, Figure 13. However, these codon groups differ significantly in the spectrum of the amino acids that they yield upon  
35 point mutation. Of all possible point mutations of one codon for serine (TCA) 78% result in a different amino acid compared to point mutations obtained for the AGT codon for serine. See, Figure 13. A virus with hundreds of codon alterations is in, statistically, a very different  
40 25 mutational space, able to access a totally different mutation spectrum, or "cloud," compared to the wild-type virus. The overall strategy for producing an evolution-defective virus is additionally set forth in Figures 14, 15 and 17. Figure 16, panels A-C show results of single  
45 mutations of different codons for ser, arg, and leu.

30 Point mutation is critical for viruses such as HIV-1 to stay ahead of the host immune system. The amino acid mutations that are required for virus escape are likely not random. Wild type codon usage has evolved to allow optimal immune system evasion. The  
50

5 wild type codon usage is likely to favor mutations that represent alterations that avoid the  
host immune system, without detrimentally affecting the protein(s) encoded. While complex,  
10 this natural pattern of amino acid sequence change of the natural virus in response to the host  
system is non-random and weakly predictable. *See also*, Seiller-Moiseiwitsch et al. (1994)

5 Annu. Rev. Genet. 28:559-596.

Changing all of the codons for ser, arg and leu in the 875 aa envelope  
15 polypeptide of HIV-1 (e.g., strain MN) would affect 187 codons (22%) resulting in 561  
mutations. *See*, e.g., Figure 18, panels A-D. If all of the HIV proteins were altered, the  
number of mutations would be more than three-fold higher. The construction of such codon  
20 modified viruses is simplified by recent advances in the synthesis of long DNA sequences,  
which enable the assembly of a plasmid of average size from 40 mer oligos in a single step  
with about 75% efficiency. *See*, Stemmer (1994) Nature 370\_389-391. *See also*, Figure 19  
for a list of oligos in one application for synthesis of HIV Env. While the synthesis of the  
25 envelope gene is sufficient, synthesis of the whole HIV genome from oligos can be  
15 performed by this method.

In practice, a preferred balance of attenuation and evolution impairment is  
30 obtained by DNA shuffling (e.g., Stemmer et al. (1995) Gene 164:49-53), e.g., of the wild-  
type and codon altered sequences, followed by selection of the resulting library of viruses  
that retain moderate growth despite many codon alterations.

20 While attenuation that can be obtained by this approach may be sufficient for  
obtaining a vaccine for most viruses, for HIV-1, the evolution impairment is more important,  
35 due to the high mutation rate of the virus. Live vaccines are used only if they elicit an  
immune response which is complex and strong enough to prevent infection of the wild-type  
virus. Live virus vaccines are typically more protective than single protein vaccines because  
40 25 it is harder to out-mutate T and B-cell responses to a larger number of epitopes. The weak  
growth of the live virus vaccine results in a larger antigenic dose and point mutation is  
increases the complexity of the immune response. To evaluate vaccine competence, vaccine  
45 potential is evaluated in Macaques (*M. nemestrina*) or chimpanzees using SIV variants that  
are known to cause AIDS. Sequence for an example SIV, SIVsmm, is found at Gene Bank  
30 Accession No. x14307. This virus is closely related to HIV-2. *See*, Hirsch (1989) Nature  
339: 389-392. In general, many complete sequences for HIVs, SIVs and many other viruses

are found in well known sequence repositories, including GenBank, EMBL, DDBJ and the NCBI. Well characterized HIV clones include: HIV-1NL43, HIV-1SF2, HIV-1BRU and HIV-1MN. For an introduction to the genetic variability of HIV, *see*, Seillier-Moiseiwitsch *et al.* (1994) Annu. Rev. Genet. 28:559-96 and the references cited therein.

Several HIV-2 isolates, including three molecular clones of HIV-2 (HIV-2<sub>ROD</sub>, HIV-2<sub>SBL-1SY</sub>, and HIV-2<sub>UC1</sub>), have also been reported to infect macaques (*M. mulatta* and *M. nemestrina*) or baboons (Franchini, *et al.* (1989) *Proc. Natl. Acad. Sci. U.S.A.* 86, 2433-2437; Barnett, *et al.* (1993) *Journal of Virology* 67, 1006-14; Boeri, *et al.* (1992) *Journal of Virology* 66, 4546-50; Castro, *et al.* (1991) *Virology* 184, 219-26; Franchini, *et al.* (1990) *Journal of Virology* 64, 4462-7; Putkonen, *et al.* (1990) *Aids* 4, 783-9; Putkonen, *et al.* (1991) *Nature* 352, 436-8). As human pathogens capable of infection of small primates, HIV-2 molecular clones provide attractive models for studies of AIDS pathogenesis, and for drug and vaccine development against HIV-1 and HIV-2.

Recently, HIV-2 was suggested as a possible vaccine candidate against the more virulent HIV-1 due to its long asymptomatic latency period, and its ability to protect against infection by HIV-1 (*see*, Travers *et al.* (1995) *Science* 268: 1612-1615 and related commentary by Cohen *et al.* (1995) *Science* 268: 1566). In the nine-year study by Travers *et al.* (*id*) of West African prostitutes infected with HIV-2, it was determined that infection with HIV-2 caused a 70% reduction in infection by HIV-1. Thus, codon altered HIV-2 viruses can also be used as a live vaccine, against both HIV-2 and HIV-1. Furthermore, because the natural pathogenicity of HIV-2 is less than HIV-1, it is, in addition to HIV-1, a preferred virus for modification.

#### FORMATS FOR SEQUENCE RECOMBINATION

The methods of the invention entail performing recombination ("shuffling") and screening or selection to "evolve" individual genes, whole plasmids or viruses, multigene clusters, or even whole genomes (Stemmer (1995) *Bio/Technology* 13:549-553). Reiterative cycles of recombination and screening/selection can be performed to further evolve the nucleic acids of interest. Such techniques do not require the extensive analysis and computation required by conventional methods for polypeptide engineering. Shuffling allows the recombination of large numbers of mutations in a minimum number of selection cycles, in contrast to natural pair-wise recombination events (e.g., as occur during sexual

5 replication). Thus, the sequence recombination techniques described herein provide particular advantages in that they provide recombination between mutations in any or all of these, thereby providing a very fast way of exploring the manner in which different  
10 combinations of mutations can affect a desired result. In some instances, however, structural and/or functional information is available which, although not required for sequence recombination, provides opportunities for modification of the technique.

15 Exemplary formats and examples for sequence recombination, referred to, e.g., as "DNA shuffling," "fast forced evolution," or "molecular breeding," have been described by the present inventors and co-workers in the following patents and patent  
20 applications: US Patent No. 5,605,793; PCT Application WO 95/22625 (Serial No. PCT/US95/02126), filed February 17, 1995; US Serial No. 08/425,684, filed April 18, 1995; US Serial No. 08/621,430, filed March 25, 1996; PCT Application WO 97/20078 (Serial No. PCT/US96/05480), filed April 18, 1996; PCT Application WO 97/35966, filed March 20, 1997; US Serial No. 08/675,502, filed July 3, 1996; US Serial No. 08/721, 824, filed  
25 September 27, 1996; PCT Application WO 98/13487, filed September 26, 1997; "Evolution of Whole Cells and Organisms by Recursive Sequence Recombination" Attorney Docket No. 018097-020720US filed July 15, 1998 by del Cardayre et al. (PCT/US99/15972, filed  
30 07/15/1999); Stemmer, *Science* 270:1510 (1995); Stemmer et al., *Gene* 164:49-53 (1995); Stemmer, *Bio/Technology* 13:549-553 (1995); Stemmer, *Proc. Natl. Acad. Sci. U.S.A.* 91:10747-10751 (1994); Stemmer, *Nature* 370:389-391 (1994); Cramer et al., *Nature Medicine* 2(1):1-3 (1996); and Cramer et al., *Nature Biotechnology* 14:315-319 (1996), each of which is incorporated by reference in its entirety for all purposes.

35 The recombination procedure starts with at least two substrates that generally show substantial sequence identity to each other (e.g., at least about 30%, 50%, 70%, 80% or  
40 90% or more sequence identity), but differ from each other at certain positions. For example, at least one codon altered nucleic acid is recombined with one or more additional nucleic acid (the additional nucleic acid can also be a codon altered nucleic acid) herein. The difference  
45 between nucleic acids to be recombined can be any type of mutation, for example, substitutions, insertions and deletions. Often, different segments differ from each other in  
30 about 5-20 positions. For recombination to generate increased diversity relative to the starting materials, the starting materials must differ from each other in at least two nucleotide

5 positions. That is, if there are only two substrates, there should be at least two divergent  
positions. If there are three substrates, for example, one substrate can differ from the second  
at a single position, and the second can differ from the third at a different single position.  
10 The starting DNA segments can be natural variants of each other, for example, allelic or  
5 species variants. More typically, they will be codon altered nucleic acids derived from one or  
more homologous nucleic acid sequence. The segments can also be from nonallelic genes  
15 showing some degree of structural and usually functional relatedness (*e.g.*, codon altered  
nucleic acids derived from different, but homologous, genes within a superfamily). The  
starting DNA segments can also be induced variants of each other. For example, one DNA  
20 segment can be produced by error-prone PCR replication of the other, or by substitution of a  
mutagenic cassette. Induced mutants can also be prepared by propagating one (or both) of  
the segments in a mutagenic strain. In these situations, strictly speaking, the second DNA  
segment is not a single segment but a large family of related segments. The different  
25 segments forming the starting materials are often the same length or substantially the same  
length. However, this need not be the case; for example; one segment can be a subsequence  
15 of another. The segments can be present as part of larger molecules, such as vectors, or can  
be in isolated form.

30 The starting DNA segments are recombined by any of the sequence  
recombination formats provided herein to generate a diverse library of recombinant DNA  
20 segments. Such a library can vary widely in size from having fewer than 10 to more than  
10<sup>5</sup>, 10<sup>9</sup>, 10<sup>12</sup>, 10<sup>15</sup>, 10<sup>20</sup> or even more members. In some embodiments, the starting  
35 segments and the recombinant libraries generated will include essentially full-length coding  
sequences and any essential regulatory sequences, such as a promoter and polyadenylation  
sequence, required for expression. In other embodiments, the recombinant DNA segments in  
40 25 the library can be inserted into a common vector providing sequences necessary for  
expression before performing screening/selection.

#### Use of Restriction Enzyme Sites to Recombine Mutations

45 In some situations it is advantageous to use restriction enzyme sites in nucleic  
acids to direct the recombination of mutations in a nucleic acid sequence of interest. These  
30 techniques are particularly preferred in the evolution of fragments that cannot readily be  
shuffled by other existing methods due to the presence of repeated DNA or other problematic  
50

5 primary sequence motifs. These situations also include recombination formats in which it is preferred to retain certain sequences unmutated. The use of restriction enzyme sites is also preferred for shuffling large fragments (typically greater than 10 kb), such as gene clusters  
10 that cannot be readily shuffled and "PCR-amplified" because of their size. Although  
5 fragments up to 50 kb have been reported to be amplified by PCR (Barnes, *Proc. Natl. Acad. Sci. U.S.A.* 91:2216-2220 (1994)), it can be problematic for fragments over 10 kb, and thus  
15 alternative methods for shuffling in the range of 10 - 50 kb and beyond are preferred. Preferably, the restriction endonucleases used are of the Class II type (Sambrook, Ausubel and Berger, *supra*) and of these, preferably those which generate nonpalindromic sticky end  
20 overhangs such as AlwI, Sfi I or BstXI. These enzymes generate nonpalindromic ends that allow for efficient ordered reassembly with DNA ligase. Typically, restriction enzyme (or  
endonuclease) sites are identified by conventional restriction enzyme mapping techniques (Sambrook, Ausubel, and Berger, *supra*), by analysis of sequence information for that gene,  
25 or by introduction of desired restriction sites into a nucleic acid sequence by synthesis (*i.e.* by  
15 incorporation of silent mutations). For example, one or more codon-altered nucleic acid can be recombined at restriction sites, e.g., with one or more nucleic acid of interest (including,  
e.g. a gene or gene cluster to be modified by recombination with the codon-altered nucleic  
30 acid).

The DNA substrate molecules to be digested can either be from *in vivo*  
20 replicated DNA, such as a plasmid preparation, or from synthetic or e.g., PCR amplified  
35 nucleic acid fragments harboring the restriction enzyme recognition sites of interest, preferably near the ends of the fragment. Typically, at least two variants of a gene of interest, each having one or more mutations, and at least one of which incorporating codon-  
40 modifications, are digested with at least one restriction enzyme determined to cut within the  
25 nucleic acid sequence of interest. The restriction fragments are then joined with DNA ligase to generate full length genes having shuffled regions. The number of regions shuffled will  
depend on the number of cuts within the nucleic acid sequence of interest. The shuffled  
45 molecules can be introduced into cells as described above and screened or selected for a desired property as described herein. Nucleic acid can then be isolated from pools (libraries),  
30 or clones having desired properties and subjected to the same procedure until a desired degree of improvement is obtained.

5 In some embodiments, at least one DNA substrate molecule or fragment thereof is isolated and subjected to mutagenesis. In some embodiments, the pool or library of  
10 religated restriction fragments are subjected to mutagenesis or additional recombination protocols before the digestion-ligation process is repeated. "Mutagenesis" as used herein  
5 comprises such techniques known in the art as PCR mutagenesis, oligonucleotide-directed mutagenesis, site-directed mutagenesis, etc., and recursive sequence recombination by any of  
15 the techniques described herein.

#### Reassembly PCR

A further technique for recombining mutations in a nucleic acid sequence  
10 utilizes "reassembly PCR." This method can be used to assemble multiple segments that have been separately evolved into a full length nucleic acid template such as a gene. This  
20 technique is performed when a pool of advantageous mutants is known from previous work or has been identified by screening mutants that may have been created by any mutagenesis  
25 technique known in the art, such as PCR mutagenesis, cassette mutagenesis, doped oligo mutagenesis, chemical mutagenesis, or propagation of the DNA template *in vivo* in mutator  
15 strains. Boundaries defining segments of a nucleic acid sequence of interest preferably lie in intergenic regions, introns, or areas of a gene not likely to have mutations of interest.  
30 Preferably, oligonucleotide primers (oligos) are synthesized for PCR amplification of segments of the nucleic acid sequence of interest, such that the sequences of the  
20 oligonucleotides overlap the junctions of two segments. The overlap region is typically about  
35 10 to 100 nucleotides in length.

Each of the segments is amplified with a set of such primers. The PCR  
products are then "reassembled" according to assembly protocols such as those discussed  
40 herein to assemble randomly fragmented genes. In brief, in an assembly protocol the PCR  
25 products are first purified away from the primers, by, for example, gel electrophoresis or size exclusion chromatography. Purified products are mixed together and subjected to about 1-10  
45 cycles of denaturing, reannealing, and extension in the presence of polymerase and deoxynucleoside triphosphates (dNTPs) and appropriate buffer salts in the absence of  
30 additional primers ("self-priming"). Subsequent PCR with primers flanking the gene are used  
50 to amplify the yield of the fully reassembled and shuffled genes. In some embodiments, the resulting reassembled genes are subjected to mutagenesis before the process is repeated.



5 In the present invention, oligos such as PCR primers can include codon  
modifications as compared to a starting sequence. In addition, oligonucleotides can form the  
basis for PCR concatemerization reactions in which overlapping hybridized oligonucleotides  
10 are extended in one or more PCR amplification cycles. In this embodiment, a template  
5 nucleic acid is not required (although a template or fragments thereof can be added to the  
amplification mixture, which can aid in the eventual reassembly of a full-length gene).  
Further details regarding oligonucleotide gene reassembly methods are found, e.g., in  
15 Cramer et al. "OLIGONUCLEOTIDE MEDIATED NUCLEIC ACID RECOMBINATION"  
filed February 5, 1999, USSN 60/118,813 and Cramer et al. "OLIGONUCLEOTIDE  
10 MEDIATED NUCLEIC ACID RECOMBINATION" filed June 24, 1999, USSN 60/141,049.

20 In a further embodiment, the PCR primers for amplification of segments of a  
nucleic acid sequence of interest are used to introduce variation into the gene of interest as  
follows. Mutations at sites of interest in a nucleic acid sequence are identified by screening  
25 or selection, by sequencing homologues of the nucleic acid sequence, and so on.  
15 Oligonucleotide PCR primers are synthesized which encode wild type or mutant information  
at sites of interest. These primers are then used in PCR mutagenesis to generate libraries of  
full length genes encoding permutations of wild type and mutant information at the  
30 designated positions. This technique is typically advantageous in cases where the screening  
or selection process is expensive, cumbersome, or impractical relative to the cost of  
20 sequencing the genes of mutants of interest and synthesizing mutagenic oligonucleotides.

35 Site Directed Mutagenesis (SDM) with Oligonucleotides Encoding  
Homologue Mutations Followed by Shuffling

In some embodiments of the invention, sequence information from one or  
more substrate sequences is added to a given "parental" sequence of interest, with subsequent  
40 25 recombination between rounds of screening or selection. Typically, this is done with site-  
directed mutagenesis performed by techniques well known in the art (e.g., Berger, Ausubel  
and Sambrook, *supra.*) with one substrate as a template and oligonucleotides encoding single  
or multiple mutations from other substrate sequences, e.g. homologous genes. After  
45 screening or selection for an improved phenotype of interest, the selected recombinant(s) can  
30 be further evolved using recursive techniques. After screening or selection, site-directed  
mutagenesis can be done again with another collection of oligonucleotides encoding

homologue mutations, and the above process repeated until the desired properties are obtained.

When the difference between two homologues is one or more single point mutations in a codon, degenerate oligonucleotides can be used that encode the sequences in both homologues. One oligonucleotide can include many such degenerate codons and still allow one to exhaustively search all permutations over that block of sequence.

When the homologue sequence space is very large, it can be advantageous to restrict the search to certain variants. Thus, for example, computer modeling tools (Lathrop *et al.* (1996) *J. Mol. Biol.*, 255: 641-665) can be used to model each homologue mutation onto the target protein and discard any mutations that are predicted to grossly disrupt structure and function. In silico genetic algorithm operations for generating and predicting mutational events are found in Selifonov and Stemmer "METHODS FOR MAKING CHARACTER STRINGS, POLYNUCLEOTIDES & POLYPEPTIDES HAVING DESIRED CHARACTERISTICS" filed 02/05/1999, USSN 60/118854.

Oligonucleotide and in silico shuffling formats

As mentioned above, at least two additional related formats are useful in the practice of the present invention. The first, referred to as "in silico" shuffling utilizes computer algorithms to perform "virtual" shuffling using genetic operators in a computer. As applied to the present invention, codon altered gene sequence strings are recombined in a computer system and desirable products are made, e.g., by reassembly PCR or ligation of synthetic oligonucleotides. In silico shuffling is described in detail in Selifonov and Stemmer in "METHODS FOR MAKING CHARACTER STRINGS, POLYNUCLEOTIDES & POLYPEPTIDES HAVING DESIRED CHARACTERISTICS" filed 02/05/1999, USSN 60/118854. In brief, genetic operators (algorithms which represent given genetic events such as point mutations, recombination of two strands of homologous nucleic acids, etc.) are used to model recombinational or mutational events which can occur in one or more nucleic acid, e.g., by aligning nucleic acid sequence strings (using standard alignment software, or by manual inspection and alignment) and predicting recombinational outcomes. The predicted recombinational outcomes are used to produce corresponding molecules, e.g., by oligonucleotide synthesis and reassembly PCR.

5 The second useful format is referred to as "oligonucleotide mediated  
shuffling" in which oligonucleotides corresponding to a family of related homologous nucleic  
acids (e.g., as applied to the present invention, codon modified synthetic homologous variants  
10 of a nucleic acid) which are recombined to produce selectable nucleic acids. This format is  
described in detail in Cramer et al. "OLIGONUCLEOTIDE MEDIATED NUCLEIC ACID  
RECOMBINATION" filed February 5, 1999, USSN 60/118,813 and Cramer et al.  
15 "OLIGONUCLEOTIDE MEDIATED NUCLEIC ACID RECOMBINATION" filed June 24,  
1999, USSN 60/141,049. In brief, selected oligonucleotides are synthesized, ligated and  
elongated, typically either in a polymerase or ligase-mediated elongation reaction. The  
10 technique can be used to recombine homologous or even non-homologous codon-altered  
nucleic acid sequences.

One advantage of oligonucleotide-mediated recombination is the ability to  
recombine homologous nucleic acids with low sequence similarity, or even non-homologous  
25 nucleic acids. In these low-homology oligonucleotide shuffling methods, one or more set of  
fragmented nucleic acids (e.g., cleaved codon-modified oligonucleotides, or synthesized  
15 codon-modified oligonucleotides) are recombined, e.g., with a set of crossover family  
diversity oligonucleotides. Each of these crossover oligonucleotides have a plurality of  
sequence diversity domains corresponding to a plurality of sequence diversity domains from  
30 homologous or non-homologous nucleic acids with low sequence similarity. The fragmented  
oligonucleotides, which are derived by comparison to one or more homologous or non-  
20 homologous nucleic acids, can hybridize to one or more region of the crossover oligos,  
35 facilitating recombination.

When recombining homologous nucleic acids, sets of overlapping family gene  
shuffling oligonucleotides (which are derived by comparison of homologous nucleic acids  
40 that include one or more codon-modified nucleic acid, followed by synthesis of  
corresponding oligonucleotides) are hybridized and elongated (e.g., by reassembly PCR or  
ligation), providing a population of recombined nucleic acids, which can be selected for a  
45 desired trait or property. The set of overlapping family shuffling gene oligonucleotides  
includes a plurality of oligonucleotide member types which have consensus region  
30 subsequences derived from a plurality of homologous target nucleic acids.

Typically, as applied to the present invention, family gene shuffling oligonucleotide which include one or more codon-altered nucleic acid(s) are provided by aligning homologous nucleic acid sequences to select conserved regions of sequence identity and regions of sequence diversity. A plurality of family gene shuffling oligonucleotides are synthesized (serially or in parallel) which correspond to at least one region of sequence diversity.

Sets of fragments, or subsets of fragments used in oligonucleotide shuffling approaches can be provided by cleaving one or more homologous nucleic acids (e.g., with a DNase), or, more commonly, by synthesizing a set of oligonucleotides corresponding to a plurality of regions of at least one nucleic acid (typically oligonucleotides corresponding to a full-length nucleic acid are provided as members of a set of nucleic acid fragments). In the shuffling procedures herein, these cleavage fragments can be used in conjunction with family gene shuffling oligonucleotides, e.g., in one or more recombination reaction to produce recombinant codon-altered nucleic acid(s).

Additional oligonucleotide shuffling formats are found in co-filed application by Cramer et al., "OLIGONUCLEOTIDE MEDIATED NUCLEIC ACID RECOMBINATION" (Attorney Docket Number 02-296-2US) and in co-filed application by Welch et al., "USE OF CODON VARIED OLIGONUCLEOTIDE SYNTHESIS FOR SYNTHETIC SHUFFLING" (Attorney docket number 02-1007). In particular, these applications provide for tri-nucleotide-based synthesis of degenerate oligonucleotides, thereby providing for codon substitution during oligonucleotide shuffling. In brief, this procedure utilizes tri-nucleotide phosphoramidite chemistry to synthesize oligos, rather than standard mono-nucleotide synthesis. Because codons are altered as a unit, the synthetic scheme of degenerate oligonucleotides is simplified.

#### Additional In Vitro DNA Shuffling Formats

In one embodiment for shuffling DNA sequences *in vitro*, the initial substrates for recombination are a pool of related sequences, e.g., different variant forms, as homologs from different individuals, strains, or species of an organism, or related sequences from the same organism, as allelic variations. The sequences can be DNA or RNA and can be of various lengths depending on the size of the gene or DNA fragment to be recombined or reassembled. Preferably the sequences are from 50 base pairs (bp) to 50 kilobases (kb).

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The pool of related substrates are converted into overlapping fragments, *e.g.*, from about 5 bp to 5 kb or more. Often, for example, the size of the fragments is from about 10 bp to 1000 bp, and sometimes the size of the DNA fragments is from about 100 bp to 500 bp. The conversion can be effected by a number of different methods, such as DNase I or RNase digestion, random shearing or partial restriction enzyme digestion, or by oligonucleotide synthesis as in the family oligonucleotide-mediated shuffling methods of crameri et al., discusses *supra*. For discussions of protocols for the isolation, manipulation, enzymatic digestion, and the like, of nucleic acids, *see*, for example, Sambrook *et al.* and Ausubel, both *supra*. The concentration of nucleic acid fragments of a particular length and sequence is often less than 0.1 % or 1% by weight of the total nucleic acid. The number of different specific nucleic acid fragments in the mixture is usually at least about 2, 10, 100, 500 or 1,000 or more.

The mixed population of nucleic acid fragments are converted to at least partially single-stranded form using any of a variety of techniques, including, for example, heating, chemical denaturation, use of DNA binding proteins, and the like (in oligonucleotide mediated methods, this step can be omitted). Conversion can be effected by heating to about 80 °C to 100 °C, more preferably from 90 °C to 96 °C, to form single-stranded nucleic acid fragments and then reannealing. Conversion can also be effected by treatment with a single-stranded DNA binding protein (see Wold (1997) *Annu. Rev. Biochem.* 66:61-92) or *recA* protein (see, *e.g.*, Kiianitsa (1997) *Proc. Natl. Acad. Sci. U S A* 94:7837-7840). Single-stranded nucleic acid fragments having regions of sequence identity with other single-stranded nucleic acid fragments can then be reannealed by cooling to 20 °C to 75 °C, and preferably from 40 °C to 65 °C. Renaturation can be accelerated by the addition of polyethylene glycol (PEG), other volume-excluding reagents or salt. The salt concentration is preferably from 0 mM to 200 mM, more preferably the salt concentration is from 10 mM to 100 mM. The salt may be KCl or NaCl. The concentration of PEG is preferably from 0% to 20%, more preferably from 5% to 10%. The fragments that reanneal can be from different substrates. The annealed nucleic acid fragments are incubated in the presence of a nucleic acid polymerase, such as Taq or Klenow, and dNTP's (*i.e.* dATP, dCTP, dGTP and dTTP). If regions of sequence identity are large, Taq polymerase can be used with an annealing temperature of between 45-65 °C. If the areas of identity are small, Klenow polymerase can

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5 be used with an annealing temperature of between 20-30 °C. The polymerase can be added to the random nucleic acid fragments prior to annealing, simultaneously with annealing or after annealing.

10 The process of denaturation, renaturation and incubation in the presence of  
5 polymerase or ligase of overlapping fragments to generate a collection of polynucleotides containing different permutations of fragments is sometimes referred to as shuffling of the nucleic acid *in vitro*. This cycle is repeated for a desired number of times. Preferably the  
15 cycle is repeated from 2 to 100 times, more preferably the sequence is repeated from 10 to 40 times. The resulting nucleic acids are a family of double-stranded polynucleotides of from  
20 about 50 bp to about 100 kb, preferably from 500 bp to 50 kb. The population represents variants of the starting substrates showing substantial sequence identity thereto but also diverging at several positions. The population has many more members than the starting substrates. The population of fragments resulting from shuffling is used to transform host  
25 cells, optionally after cloning into a vector.

15 In one embodiment utilizing *in vitro* shuffling, subsequences of recombination substrates can be generated by amplifying the full-length sequences under conditions which produce a substantial fraction, typically at least 20 percent or more, of incompletely extended  
30 amplification products. Another embodiment uses random primers to prime an entire template DNA to generate less than full length amplification products. The amplification  
20 products, including the incompletely extended amplification products are denatured and subjected to at least one additional cycle of reannealing and amplification. This variation, in  
35 which at least one cycle of reannealing and amplification provides a substantial fraction of incompletely extended products, is termed "stuttering." In the subsequent amplification round, the partially extended (less than full length) products reanneal to and prime extension  
40 25 on different sequence-related template species. In another embodiment, the conversion of substrates to fragments can be effected by partial PCR amplification of substrates.

In another embodiment, a mixture of fragments is spiked with one or more  
45 oligonucleotides. The oligonucleotides can be designed to include precharacterized mutations of a wildtype sequence (e.g., codon modification), or sites of natural variations  
30 between individuals or species. The oligonucleotides also typically include sufficient sequence or structural homology flanking such mutations or variations to allow annealing  
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5 with the wildtype fragments. Annealing temperatures can be adjusted depending on the length of homology.

10 In a further embodiment, recombination occurs in at least one cycle by template switching, such as when a DNA fragment derived from one template primes on the homologous position of a related but different template. Template switching can be induced  
5 by addition of rccA (*see*, Kiianitsa (1997) *supra*), rad51 (*see*, Namsaraev (1997) *Mol. Cell. Biol.* 17:5359-5368), rad55 (*see*, Clever (1997) *EMBO J.* 16:2535-2544), rad57 (*see*, Sung (1997) *Genes Dev.* 11:1111-1121) or other polymerases (*e.g.*, viral polymerases, reverse transcriptase) to the amplification mixture. Template switching can also be increased by  
15 increasing the DNA template concentration.

20 Another embodiment utilizes at least one cycle of amplification, which can be conducted using a collection of overlapping single-stranded DNA fragments of related sequence, and different lengths. Fragments can be prepared using a single stranded DNA phage, such as M13 (*see*, Wang (1997) *Biochemistry* 36:9486-9492). Each fragment can  
25 hybridize to and prime polynucleotide chain extension of a second fragment from the collection, thus forming sequence-recombined polynucleotides. In a further variation, ssDNA fragments of variable length can be generated from a single primer by Pfu, Taq, Vent, Deep Vent, UITma DNA polymerase or other DNA polymerases on a first DNA template  
30 (*see*, Cline (1996) *Nucleic Acids Res.* 24:3546-3551). The single stranded DNA fragments are used as primers for a second, Kunkel-type template, consisting of a uracil-containing circular ssDNA. This results in multiple substitutions of the first template into the second.  
35 *See*, Levichkin (1995) *Mol. Biology* 29:572-577; Jung (1992) *Gene* 121:17-24.

In some embodiments of the invention, shuffled nucleic acids obtained by use of the recursive recombination methods of the invention, are put into a cell and/or organism  
40 25 for screening. Shuffled genes can be introduced into, for example, bacterial cells, yeast cells, fungal cells vertebrate cells, invertebrate cells or plant cells for initial screening. *Bacillus* species (such as *B. subtilis* and *E. coli* are two examples of suitable bacterial cells into which one can insert and express shuffled genes which provide for convenient shuttling to other cell  
45 types (a variety of vectors for shuttling material between these bacterial cells and eukaryotic cells are available; *see*, Sambrook, Ausubel and Berger, *all supra*). The shuffled genes can  
30

5 be introduced into bacterial, fungal or yeast cells either by integration into the chromosomal  
DNA or as plasmids.

10 Bacterial, plant, animal and yeast systems are preferred in the present  
invention. For example, in one embodiment, shuffled genes can be introduced into plant or  
5 animal cells for production purposes (it will be appreciated that transgenic plants are,  
increasingly, an important source of industrial enzymes), or can be introduced into a plant or  
15 animal cell for therapeutic purposes. Thus, a transgene of interest can be modified using the  
recursive sequence recombination methods of the invention *in vitro* and reinserted into the  
cell for *in vivo/in situ* selection for the new or improved property, in bacteria, eukaryotic  
10 cells, or whole eukaryotic organisms.

#### 20 In Vivo DNA Shuffling Formats

In some embodiments of the invention, DNA substrate molecules, e.g., those  
comprising codon modifications relative to a wild-type sequence, are introduced into cells,  
25 where the cellular machinery directs their recombination. For example, a library of mutants  
15 is constructed and screened or selected for mutants with improved phenotypes by any of the  
techniques described herein.

30 The DNA substrate molecules encoding the best candidates are recovered by  
any of the techniques described herein, then fragmented and used to transfect a plant host and  
screened or selected for improved function. If further improvement is desired, the DNA  
20 substrate molecules are recovered from the host cell, such as by PCR, and the process is  
35 repeated until a desired level of improvement is obtained. In some embodiments, the  
fragments are denatured and reannealed prior to transfection, coated with recombination  
stimulating proteins such as recA, or co-transfected with a selectable marker such as Neo<sup>R</sup> to  
allow the positive selection for cells receiving recombined versions of the gene of interest.  
40 25 Methods for *in vivo* shuffling are described in, for example, PCT application WO 98/13487  
and WO 97/20078. The efficiency of *in vivo* shuffling can be enhanced by increasing the  
copy number of a gene of interest in the host cells.

#### 45 Whole Genome Shuffling

In one embodiment, the selection methods herein are utilized in a "whole  
30 genome shuffling" format. An extensive guide to the many forms of whole genome shuffling  
is found in the pioneering application to the inventors and their co-workers entitled  
50



5 "Evolution of Whole Cells and Organisms by Recursive Sequence Recombination,"  
PCT/US99/15972, by del Cardayre et al. Any codon-altered set of nucleic acids can be used  
10 to transform cells, which can then be shuffled by in a whole genome format.

15 In brief, whole genome shuffling makes no presuppositions at all regarding  
5 what nucleic acids may confer a desired property. Instead, entire genomes (e.g., from a  
genomic library, or isolated from an organism) are shuffled in cells and selection protocols  
15 applied to the cells. These genomes can be spiked with any desired set of nucleic acids,  
including codon-modified nucleic acids.

#### Assays

20 The relevant assay for selection of a desired property of a codon-modified  
nucleic acid will depend on the application. Many assays which detect activity for proteins,  
receptors, ligands, cells and the like are known. Formats include binding to immobilized  
components, cell or organismal viability, production of reporter compositions, and the like.

25 In the high throughput assays of the invention, it is possible to screen up to  
15 several thousand different shuffled variants in a single day. In particular, each well of a  
microtiter plate can be used to run a separate assay, or, if concentration or incubation time  
effects are to be observed, every 5-10 wells can test a single variant. Thus, a single standard  
30 microtiter plate can assay about 100 (e.g., 96) reactions. If 1536 well plates are used, then a  
single plate can easily assay from about 100- about 1500 different reactions. It is possible to  
20 assay several different plates per day; assay screens for up to about 6,000-20,000 different  
assays (i.e., involving different nucleic acids, encoded proteins, concentrations, etc.) is  
35 possible using the integrated systems of the invention. More recently, microfluidic  
approaches to reagent manipulation have been developed, e.g., by Caliper Technologies  
(Mountain View, CA).

40 25 In one aspect, library members, e.g., cells, viral plaques, spores or the like, are  
separated on solid media to produce individual colonies (or plaques). Using an automated  
colony picker (e.g., the Q-bot, Genetix, U.K.), colonies or plaques are identified, picked, and  
45 up to 10,000 different mutants inoculated into 96 well microtitre dishes containing two 3 mm  
glass balls/well. The Q-bot does not pick an entire colony but rather inserts a pin through the  
30 center of the colony and exits with a small sampling of cells, (or mycelia) and spores (or  
viruses in plaque applications). The time the pin is in the colony, the number of dips to

5 inoculate the culture medium, and the time the pin is in that medium each effect inoculum size, and each can be controlled and optimized. The uniform process of the Q-bot decreases human handling error and increases the rate of establishing cultures (roughly 10,000/4 hours).

10 These cultures are then shaken in a temperature and humidity controlled incubator. The glass balls in the microtiter plates act to promote uniform aeration of cells and the dispersal of mycelial fragments similar to the blades of a fermenter. Clones from cultures of interest can be cloned by limiting dilution. As also described supra, plaques or cells constituting libraries can also be screened directly for production of proteins, either by detecting hybridization, protein activity, protein binding to antibodies, or the like.

15 The ability to detect a subtle increase in the performance of a shuffled library member over that of a parent strain relies on the sensitivity of the assay. The chance of finding the organisms having an improvement is increased by the number of individual mutants that can be screened by the assay. To increase the chances of identifying a pool of sufficient size, a prescreen that increases the number of mutants processed by, e.g., 10-fold can be used. The goal of the primary screen is to quickly identify mutants having equal or better product titres than the parent strain(s) and to move only these mutants forward to liquid cell culture for subsequent analysis.

20 A number of well known robotic systems have also been developed for solution phase chemistries useful in assay systems. These systems include automated workstations like the automated synthesis apparatus developed by Takeda Chemical Industries, LTD. (Osaka, Japan) and many robotic systems utilizing robotic arms (Zymate II, Zymark Corporation, Hopkinton, Mass.; Orca, Hewlett-Packard, Palo Alto, Calif.) which mimic the manual synthetic operations performed by a scientist. Any of the above devices are suitable for use with the present invention, e.g., for high-throughput screening of molecules encoded by codon-altered nucleic acids. The nature and implementation of modifications to these devices (if any) so that they can operate as discussed herein with reference to the integrated system will be apparent to persons skilled in the relevant art.

25 High throughput screening systems are commercially available (*see, e.g.*, Zymark Corp., Hopkinton, MA; Air Technical Industries, Mentor, OH; Beckman Instruments, Inc. Fullerton, CA; Precision Systems, Inc., Natick, MA, *etc.*). These systems typically automate entire procedures including all sample and reagent pipetting, liquid

5 dispensing, timed incubations, and final readings of the microplate in detector(s) appropriate for the assay. These configurable systems provide high throughput and rapid start up as well as a high degree of flexibility and customization.

10 The manufacturers of such systems provide detailed protocols the various high  
5 throughput. Thus, for example, Zymark Corp. provides technical bulletins describing screening systems for detecting the modulation of gene transcription, ligand binding, and the like. Microfluidic approaches to reagent manipulation have also been developed, e.g., by  
15 Caliper Technologies (Mountain View, CA).

Optical images viewed (and, optionally, recorded) by a camera or other  
10 recording device (e.g., a photodiode and data storage device) are optionally further processed in any of the embodiments herein, e.g., by digitizing the image and/or storing and analyzing the image on a computer. A variety of commercially available peripheral equipment and software is available for digitizing, storing and analyzing a digitized video or digitized  
20 optical image, e.g., using PC (Intel x86 or pentium chip- compatible DOS™, OS2™  
25 WINDOWS™, WINDOWS NT™ or WINDOWS95™ based machines), MACINTOSH™,  
15 or UNIX based (e.g., SUN™ work station) computers.

One conventional system carries light from the assay device to a cooled  
30 charge-coupled device (CCD) camera, in common use in the art. A CCD camera includes an array of picture elements (pixels). The light from the specimen is imaged on the CCD.  
20 Particular pixels corresponding to regions of the specimen (e.g., individual hybridization sites on an array of biological polymers) are sampled to obtain light intensity readings for each  
35 position. Multiple pixels are processed in parallel to increase speed. The apparatus and methods of the invention are easily used for viewing any sample, e.g., by fluorescent or dark field microscopic techniques.

40 25 Software elements for manipulating strings of characters which correspond to codon-modified nucleic acids can be used to direct synthesis of oligonucleotides relevant to shuffling of codon-modified nucleic acids. Integrated systems comprising these and other  
45 useful features, e.g., a digital computer with additional features such as high-throughput liquid control software, image analysis software, data interpretation software, a robotic liquid  
30 control armature for transferring solutions from a source to a destination operably linked to the digital computer, an input device (e.g., a computer keyboard) for entering data to the

5 digital computer to control high throughput liquid transfer by the robotic liquid control  
armature an image scanner for digitizing label signals from labeled assay components, or the  
like are a feature of the invention.

10 In one aspect, the invention provides an integrated system comprising a  
5 computer or computer readable medium comprising a database having at least two artificial  
homologous codon-altered nucleic acid sequence strings, and a user interface allowing a user  
to selectively view one or more sequence strings in the database. As discussed throughout,  
15 there are a variety of sequence database programs for aligning and manipulating sequences.  
In addition, standard text manipulation software such as word processing software (e.g.,  
20 Microsoft Word™ or Corel Wordperfect™) and database software (e.g., spreadsheet software  
such as Microsoft Excel™, Corel Quattro Pro™, or database programs such as Microsoft  
Access™ or Paradox™) can be used in conjunction with a user interface (e.g., a GUI in a  
standard operating system such as a Windows, Macintosh or LINUX system) to manipulate  
25 strings of characters. Specialized alignment programs such as BLAST can also be  
15 incorporated into the systems of the invention for alignment of codon-altered nucleic acids  
(or corresponding character strings).

30 In addition to the integrated system elements mentioned above, the integrated  
system can also include an automated oligonucleotide synthesizer operably linked to the  
computer or computer readable medium. Typically, the synthesizer is programmed to  
20 synthesize one or more oligonucleotide comprising one or more subsequence of one or more  
of the at least two artificial homologous codon-altered nucleic acids.

35 Modifications can be made to the method and materials as hereinbefore  
described without departing from the spirit or scope of the invention as claimed, and the  
invention can be put to a number of different uses, including:

40 25 The use of an integrated system to test shuffled codon-modified DNAs,  
including in an iterative process.

45 An assay, kit or system utilizing a use of any one of the selection strategies,  
materials, components, methods or substrates hereinbefore described. Kits will optionally  
additionally comprise instructions for performing methods or assays, packaging materials,  
30 one or more containers which contain assay, device or system components, or the like.

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In an additional aspect, the present invention provides kits embodying the methods and apparatus herein. Kits of the invention optionally comprise one or more of the following: (1) a shuffled codon-modified component as described herein; (2) instructions for practicing the methods described herein, and/or for operating the selection procedure herein; (3) one or more assay component; (4) a container for holding nucleic acids or enzymes, other nucleic acids, transgenic plants, animals, cells, or the like, (5) packaging materials and (6) software fixed in a computer readable medium comprising sequences corresponding to one or more codon-altered nucleic acid character string.

20

In a further aspect, the present invention provides for the use of any component or kit herein, for the practice of any method or assay herein, and/or for the use of any apparatus or kit to practice any assay or method herein.

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While the foregoing invention has been described in some detail for purposes of clarity and understanding, it will be clear to one skilled in the art from a reading of this disclosure that various changes in form and detail can be made without departing from the true scope of the invention. For example, all the techniques and materials described above can be used in various combinations. All publications and patent documents cited in this application are incorporated by reference in their entirety for all purposes to the same extent as if each individual publication or patent document were so individually denoted.

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**Claims**

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WHAT IS CLAIMED IS:

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1. A method of making codon altered nucleic acids, the method comprising:

(i) providing a first nucleic acid sequence, which nucleic acid sequence encodes a first

5 polypeptide sequence;

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(ii) providing a plurality of codon altered nucleic acid sequences, each of which encode the first polypeptide or a modified form thereof; and,

(iii) recombining the plurality of codon-altered nucleic acid sequences to produce a target codon altered nucleic acid, which target codon altered nucleic acid encodes a second protein.

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2. The method of claim 1, wherein at least one of the plurality of codon altered nucleic acid sequences does not hybridize to the first nucleic acid under stringent hybridization conditions.

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3. The method of claim 1, further comprising shuffling a nucleic acid comprising a subsequence consisting of the first nucleic acid, or a substantially identical

15 variant thereof, with one or more of the plurality of codon altered nucleic acids, or with the target codon altered nucleic acid.

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4. The method of claim 1, the method further comprising the step of:

(iv) screening the second protein for a structural or functional property.

35

5. The method of claim 1, the method further comprising the steps of:

20 (iv) screening the second protein for a structural or functional property, and,

(v) comparing the structural or functional property of the second protein to a structural or functional property of the first protein.

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6. The method of claim 1, wherein the second polypeptide has a structural or functional property equivalent or superior to the first polypeptide.

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25 7. The method of claim 1, wherein the first and second polypeptide are homologous.

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8. The method of claim 1, wherein the plurality of codon altered nucleic acids comprise a library of codon altered nucleic acids.

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9. The method of claim 1, wherein the plurality of codon altered nucleic acids comprise a library of codon altered conservatively modified nucleic acids.

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10. A library of codon altered conservatively modified nucleic acids produced by the method of claim 9.

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11. The method of claim 1, wherein the plurality of codon altered nucleic acids comprise a library of codon altered non-conservatively modified nucleic acids.

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12. A library of codon altered conservatively modified nucleic acids produced by the method of claim 11.

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13. The method of claim 1, wherein the plurality of codon altered nucleic acids is derived from a plurality of forms of the first nucleic acid.

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14. The method of claim 1, wherein the plurality of codon altered nucleic acid sequences comprise at least three codon altered nucleic acids.

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15. The method of claim 1, wherein the plurality of codon altered nucleic acid sequences comprise one or more of the following structural features:

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(a) codon usage divergence for each of the codon altered nucleic acids of 50% or more as compared to the first nucleic acid;

(b) codon usage divergence for each of the codon altered nucleic acids of 75% or more as compared to the first nucleic acid;

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(c) codon usage divergence for each of the codon altered nucleic acids of 90% or more as compared to the first nucleic acid;

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(d) maximal codon usage divergence for each of the codon altered nucleic acids as compared to the first nucleic acid;

(e) non-overlapping non-conservative substitutions in each of the codon altered nucleic acids as compared to the first nucleic acid;

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(f) a lack of high stringency hybridization between one or more of the codon altered nucleic acid and the first nucleic acid; and,

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(g) modification of the codons of one or more of the codon altered nucleic acids to provide one or more different hydrophobic core residue for an encoded polypeptide as compared to the first polypeptide.

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16. The method of claim 1, wherein the percent identity between the second protein and the first protein is lower than the percent identity between two of the plurality of codon altered nucleic acids.

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17. The method of claim 1, wherein the first nucleic acid encodes a protein selected from: EPO, G-CSF, a viral envelope protein, a cytokine, and a phosphatase.

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18. The method of claim 1, wherein the first nucleic acid sequence or the codon altered nucleic acid sequences are isolated nucleic acids.

19. The target codon altered nucleic acid produced by the method of claim 1.

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20. The method of claim 1, wherein the first nucleic acid sequence or the codon altered nucleic acid sequences are nucleic acids present in cells.

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21. The cells produced by the method of claim 20.

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22. The method of claim 1, wherein each of the codon altered nucleic acid sequences comprises at least two nucleotide differences when compared to the first nucleic acid.

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23. The method of claim 1, further comprising introducing the target codon altered nucleic acid into a cell, or into a vector or virus.

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24. The cell, vector or virus produced by the method of claim 23.

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25. The method of claim 1, wherein the target codon altered nucleic acid is recombined with a portion of a viral genome to produce an attenuated virus.

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26. The attenuated virus produced by the method of claim 25.

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27. The method of claim 1, wherein the target codon altered nucleic acid is recombined with a portion of a viral genome to produce an attenuated virus, which attenuated virus produces an immune response upon infection by the virus in a mammal.

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28. The attenuated virus produced by the method of claim 27.

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29. The method of claim 1, wherein the target codon altered nucleic acid is recombined with a portion of a retroviral genome to produce an attenuated retrovirus, which attenuated retrovirus produces an immune response upon infection by the retrovirus in a mammal.

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30. The attenuated retrovirus produced by the method of claim 29.

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31. The method of claim 1, wherein the target codon altered nucleic acid is recombined with a portion of a viral genome to produce an viral vector.

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32. The viral vector produced by the method of claim 31.

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33. The method of claim 1, wherein the target codon altered nucleic acid is recombined with a portion of a viral genome to produce an viral vector, which vector requires *trans* complementation for replication, and which vector has a reduced rate of reversion to a replicative form as compared to a corresponding viral vector which lacks a subsequence corresponding to the target codon altered nucleic acid.

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34. The viral vector produced by the method of claim 33.

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35. The method of claim 33, wherein the vector comprises viral elements from one or more of: a lentivirus, an adenovirus, a herpes virus, and an adeno-associated virus.

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36. A method of making a library of codon-altered nucleic acids, the method comprising:

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(i) selecting a first nucleic acid sequence, which nucleic acid sequence encodes a first polypeptide sequence; and,

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(ii) making a plurality of codon altered nucleic acid sequences, each of which encode the first polypeptide or a modified form thereof, wherein the plurality of codon altered nucleic acids comprise the library.

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37. A codon-altered library made by the method of claim 36.

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38. The library of claim 37, wherein said library comprises at least 2 codon altered nucleic acids.

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39. The library of claim 37, wherein said library comprises at least 5 codon altered nucleic acids.

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40. The library of claim 37, wherein said library comprises at least 10 codon altered nucleic acids.

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41. The library of claim 37, wherein said library comprises at least 100 codon altered nucleic acids.

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42. A kit comprising the library of claim 25 and one or more of: a container and instructional materials providing method step instructions for recombining two or more of members of the library.

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43. The method of claim 41, further comprising recombining said plurality of codon altered nucleic acids to produce a shuffled codon-altered library.

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44. The codon altered library made by the method of claim 43.

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45. The method of claim 36, wherein said nucleic acids encode a protein selected from EPO, a cytokine, a phosphatase, and a viral envelope protein.

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46. A composition comprising a plurality of codon altered nucleic acids, each of which encode a first polypeptide or a modified form thereof.

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47. A library of codon-altered nucleic acids, comprising a plurality of codon-altered nucleic acids derived from a plurality of homologous nucleic acids.

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48. The library of claim 47, wherein said plurality of codon altered nucleic acids recombine in vitro at an increased rate compared to said plurality of homologous nucleic acids.

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49. The library of claim 47, wherein the level of identity among said plurality of codon-altered nucleic acids is at least as high as among a plurality of polypeptides encoded by said plurality of homologous nucleic acids.

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50. An integrated system comprising a computer or computer readable medium comprising a database having at least two artificial homologous codon-altered nucleic acid sequence strings, and a user interface allowing a user to selectively view one or more sequence strings in the database.

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51. The integrated system of claim 50, further comprising an automated oligonucleotide synthesizer operably linked to the computer or computer readable medium, which synthesizer is programmed to synthesize one or more oligonucleotide comprising one or more subsequence of one or more of the at least two artificial homologous codon-altered nucleic acids.

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**EPO wobble gene**

Highlevel Report of EPO degeneracy.MEG, using Clustal method with Weighted residue weight table  
 Wednesday, August 05, 1998 11:29 AM

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	10	20	30	40	50	
	G C C C C A C C A C G C C T C A T C T G T C A C A G C C C A G T C T G G A G A G G T A C C T C T T					EPO coding
	G . . . . T A . G T . G . A . . C . . T T C G A . G . . . . T . A . . A C . C . . T T . G C .					EPO wobble
	INTRON					
	60	70	80	90	100	
51	G G A G C C C A A G G A G G C C C A G A A T A T C A C G C A C G C C T G T G C T G A A C A C T G C A					EPO coding
51	C . . A . . G . . A . . A . . G . . A . . C . . A . . C . . C . . G . . C . . A . . G . . T . . T T					EPO wobble
	110	120	130	140	150	
101	G C T T C A A T G A G A A T A T C A C T G C C C C A G A C A C C A A A G T T A A T T C T A T G C C					EPO coding
101	C C C . C . . C . . A . . . . . A . . . . . A . . . . . C . . . . . T . . . . . C . . . . . G					EPO wobble
	INTRON					
	160	170	180	190	200	
151	T C G A A G A G G A T G S A T G T C G G C C A G C A G G C C G T A G A A G T C T G C C A G G G C C T					EPO coding
151	A C . C . . . . . . . . . . . C . . A . . A . . G . . T . . G . . G . . A . . G T .					EPO wobble
	210	220	230	240	250	
201	G C C C C T G C C C T T G G A A G C T G T C C T C G G G G C C A G G C C C T G T T G G T C A A C T					EPO coding
201	A . . . . . T . . A T . A A G C . . . . . A . . . . . T . . A A . A . . G . . A . . C T . A C . C . . G . . T A					EPO wobble
	260	270	280	290	300	
251	C T T C C C A G C C C T G G C C A G C C C C T G C A G C T G C A T T G G A T A A A G C C G T C A G T					EPO coding
251	G C A G T . . A . . C . . . . . A . . . . . T . . A . . A T . A . . C . . C . . C . . G . . G . . G T C A					EPO wobble
	INTRON					
	310	320	330	340	350	
301	G G C C C T C C C A G C C C T A C C A C T C T G C T T C G G G C C T C T G G A G C C C A G A A G G A					EPO coding
301	. . . . . T . . A A . C T C C T . G . . G . . A T . A T . A A . A . . A T . A . . T . G . . A . . A . .					EPO wobble
	360	370	380	390	400	
351	A G C C A T C T C C C C T C C A G A T C C G C C C T C A G C T G C T C C A C T C C G A A C A A T C A					EPO coding
351	G . . . . . A A G T . . A . . T . . C . . C . . G A G T . . A . . A . . T T . G A . G . . T . . A .					EPO wobble
	410	420	430	440	450	
401	C T C T G A C A C T T T C C G C A A A C T C T C C G A G T C T A C T C C A A T T C C T C C G G					EPO coding
401	. . A . . A . . T . . A . . T A . G . . G T . G . . T A . G . . . . . T A G T . . C . . T T . G A . A					EPO wobble
	INTRON					
	460	470	480	490	500	
451	G C A A A G C C G A A G C T G T A C A C A G G G G A G C C T G C A G G A C A G G G A C A G A T C					EPO coding
451	. . . . . T . . A T . A . . A T . T . C . . A . . G . . T C . C . . T . . C . . T C . T . A					EPO wobble
	INTRON					
501						EPO coding
501						EPO wobble

Decorations: Decoration #1: Hide (as ...) residues that match EPO coding exactly.

Fig. 2



# Human EPO Wobble Sequence Space

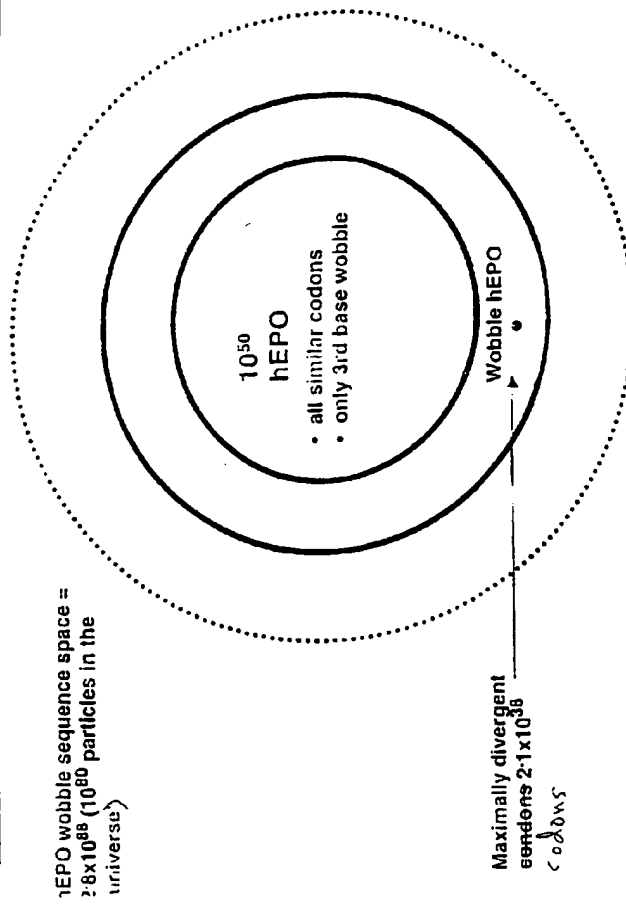
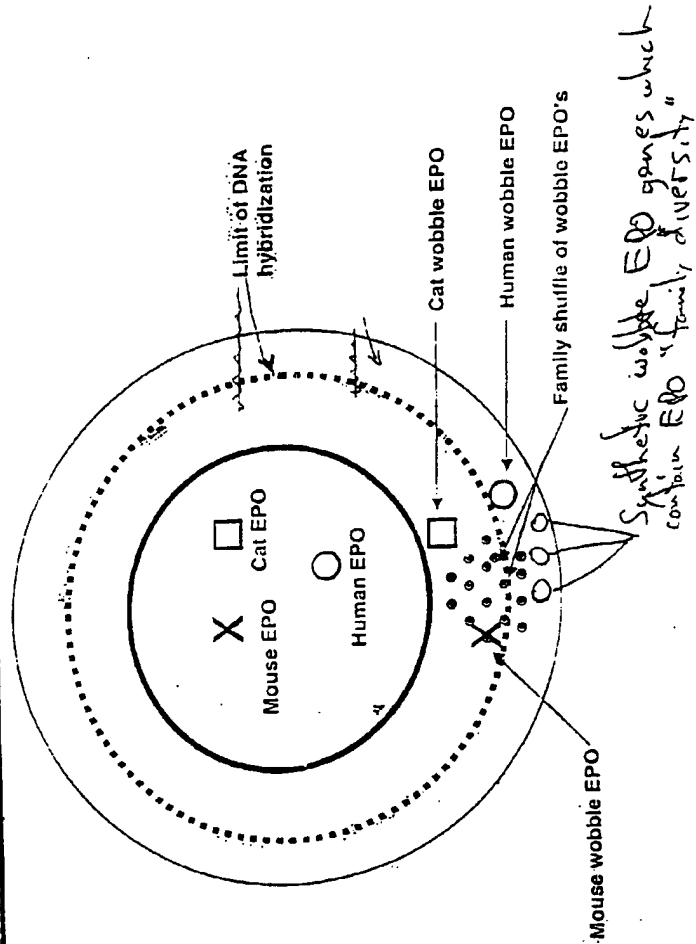


Fig. 4



Fig. 5

# Mammalian EPO Family-Wobble Sequence Space



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Current Report of goat homologs.MEG, using Chastal method with PAM250 residue weight table.  
Friday, August 08, 1998 9:08 PM

Page 1

10	20	30	40	50	Majority
TPLE-----GPASSLPQCSFLLKCLEQVRRIQADGAELOERLCATNKLCHP					human G-CSF
.....GPAFELPSSFLLRGLGMYRKTQGSAAALQSLCAATGLCHP					sheep
.....R.....A.....E.....R.....AM.....					cow
.....T.....V.....A.....T.....R.....AH.....					cat
A.....TGF.....M.....V.....A.....T.....T.....HQ.....					dog
A.....S.....A.....E.....R.....H.....					pig
V.....VTVALP.....SLP.....R.....S.....AS.....SV.....L.....Q.....					mouse
I.....LTVSSLP.....SLP.....R.....S.....ARMYK.....L.....Q.....					rat
60	70	80	90	100	Majority
RELVLGKSELCTFOAFLLSSCSALOLTGCLGQLESGLFLYQGLLQALAG					human G-CSF
.....XLVLGKSELCTFOAFLLSSCSALOLTGCLGQLESGLFLYQGLLQALAG					sheep
.....M.....R.....Q.....S.....S.....A.....N.....G.....					cow
.....A.....A.....Q.....S.....S.....T.....R.....					cat
.....A.....A.....Q.....S.....S.....M.....R.....					dog
.....Q.....L.....Q.....S.....S.....T.....H.....G.....V.....					pig
.....Q.....X.....S.....C.....S.....QTG.....C.....					mouse
.....F.....X.....S.....S.....QTE.....					rat
110	120	130	140	150	Majority
ESPELAPTLDTLQLDVEDFATNMQQHEELCHAPAVQPTQGTMTFTTSAP					human G-CSF
.....SPELAPTLDTLQLDVEDFATNMQQHEELCHAPAVQPTQGTMTFTTSAP					sheep
.....A.....X.....Y.....L.....D.....L.....D.....A.....V.....					cow
.....A.....A.....M.....T.....T.....IN.....D.....V.....P.....T.....T.....					cat
.....A.....A.....M.....T.....T.....IN.....D.....V.....P.....T.....T.....					dog
.....A.....A.....M.....T.....T.....IN.....D.....R.....SL.....TV.....T.....					pig
.....A.....A.....M.....T.....T.....IN.....D.....R.....SL.....TV.....T.....					mouse
1.....A.....A.....M.....T.....T.....IN.....D.....R.....SL.....TV.....T.....					rat
2.....S.....A.....M.....R.....DN.....S.....V.....TV.....ST.....I.....T.....					
160	170	180	190	Majority	
QRRAGGVLEVASQLQSFLEELAYRALRRLLAP-----				human G-CSF	
5QRRAGGVLEVASQLQSFLEELAYRALRRLLAP-----				sheep	
5.....Q.....R.....G.....L.....A.....C.....Y.....E.....				cow	
5.....Q.....R.....G.....L.....A.....C.....Y.....E.....				cat	
5.....T.....M.....N.....A.....A.....FTR.....				dog	
5.....M.....N.....A.....A.....FTR.....				pig	
5.....V.....Q.....L.....A.....A.....Y.....E.....				mouse	
1.....A.....E.....Y.....G.....TARDA.....E.....				rat	
1.....T.....Y.....T.....A.....H.....E.....PR.....AQKNFFESLFIS I					

covation 'covation #1': Hide (as '-') residues that match human G-CSF exactly.

Fig. 6

# G-CSF Family Diversity

TPQGPASSLPQSHLILKCIHQVVKIQGDDGAAIQKILCAI  
 A S T R P R S M V A S N T B L R T  
 V P S C L R S V Q  
 I

YKLCHEPRLV L G H S L G I P W A F L S S C P S Q A L O L A G U L  
 H Q Q M F R A L Q P S G S S O T S  
 X R Q  
 M K

SQLHSGLPLV Q G I L Q A L E Q I S F E L G P T I D T L Q L D V A D  
 D C V C A S A A A M H I T N  
 N I I T D

KATTIWQOMERL O M A P A L O P T T O G A M P A F A S A F O R R A G  
 L I N L D V R V T V P S T V T I  
 N A S L

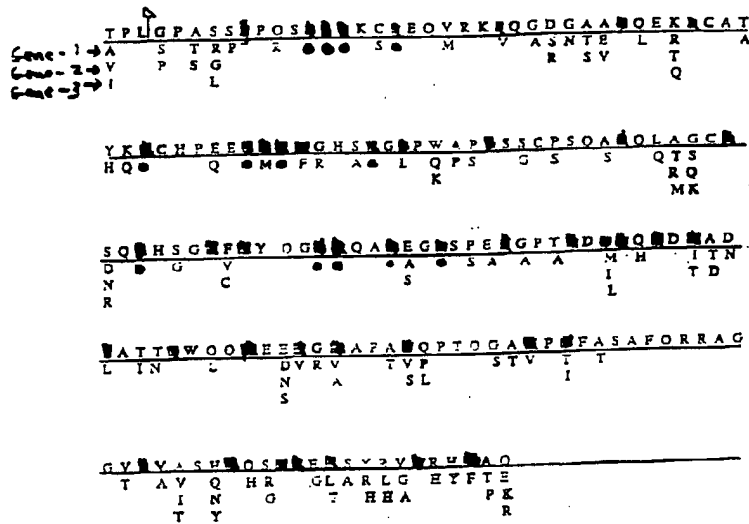
QVLVASHLQKFLVSVYRLRLA O  
 T A V Q H A G L A R L G H Y F T E  
 I N G T H H A P K  
 T Y

Fig. 7

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Fig. 8

# G-CSF G-CSG Family Diversity



• = hydrophobic core residues in conserved stretches of the protein

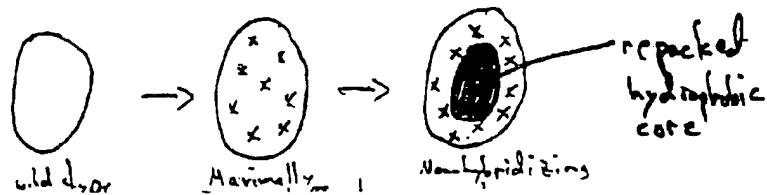
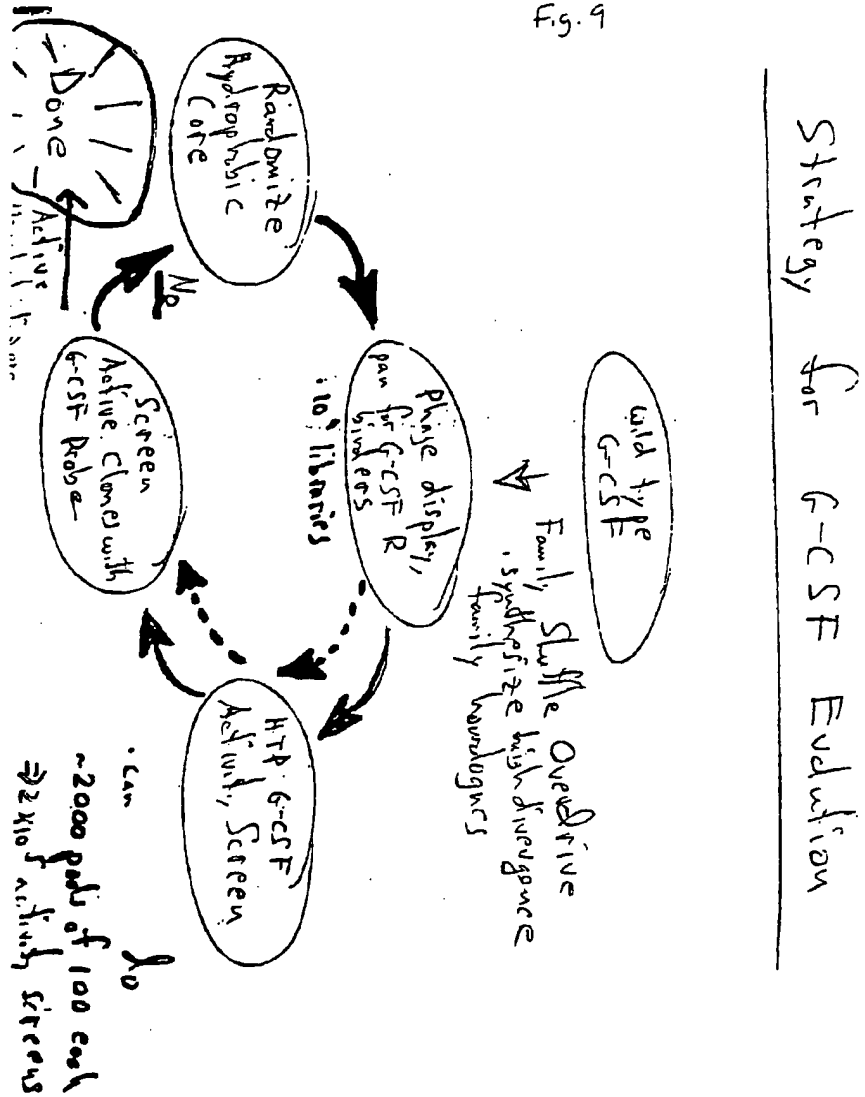


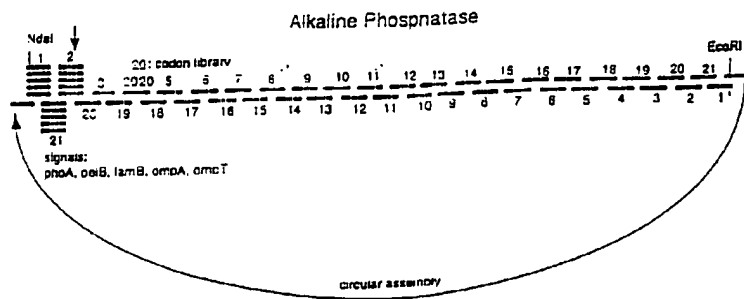
Fig. 9





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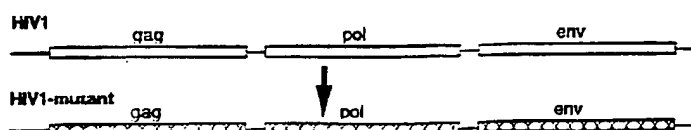
Fig. 11



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### Vaccination with evolution-defective viruses

Preventing viral immune escape by altering the mutation spectrum through massive codon usage alteration



Same current protein sequence,  
but mutations result in a different future protein sequence,  
a different and unsophisticated mutant cloud.

<p><b>Most Effective</b></p> <p>Ser : TCX ↔ AGY Arg : CGX ↔ AGR Leu : CTX ↔ TTR</p>	<p><b>Least Effective</b></p> <p><u>Phe, Tyr, Cys, His, Asp, Asn:</u> -- C ↔ -- T <u>Lys, Gln, Glu</u> -- A ↔ -- G</p>
<p><b>Moderately Effective</b></p> <p><u>Thr, Pro, Val, Ala, Gly:</u> -- R ↔ -- Y Ile : ATY ↔ ATA</p>	<p><b>Unchangeable:</b></p> <p>Trp : TGG Met : ATG</p>

Fig. 12



Ser TCX → AGY  
 Arg CGX → AGR  
 Leu CTX → TTR

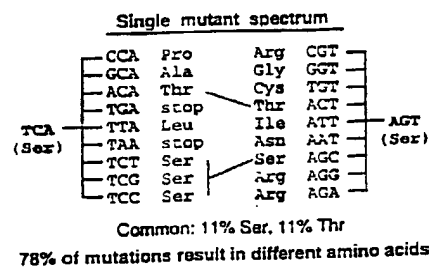


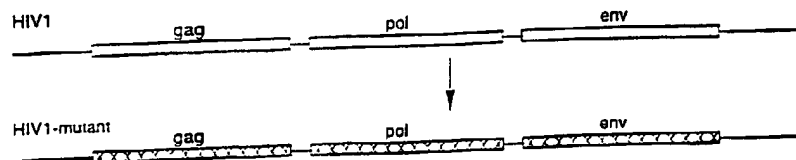
Fig. 13

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Figure 14

## Vaccination with evolution-defective viruses

Preventing viral immune escape by altering the mutation spectrum through massive codon usage alteration



Same current protein sequence,  
but mutations result in a different future protein sequence,  
a different and unsophisticated mutant cloud.

Maximizing the difference requires the following changes:

<p><b>Most Effective</b></p> <p>Ser: TCX ↔ AGY Arg: CGX ↔ AGR Leu: CTX ↔ TTR</p>	<p><b>Least Effective</b></p> <p><u>Phe, Tyr, Cys, His, Asn, Asp:</u> --C ↔ --T</p> <p><u>Lys, Gln, Glu</u> --A ↔ --G</p>
<p><b>Moderately Effective</b></p> <p><u>Thr, Pro, Val, Ala, Gly:</u> --R ↔ --Y</p> <p>Ile: ATY ↔ ATA</p>	<p><b>Unchangeable:</b></p> <p>Trp: TGG Met: ATG</p>

Fig. 15

# Vaccination with evolution-defective viruses

Preventing viral immune escape by altering the mutation spectrum through massive codon usage alteration

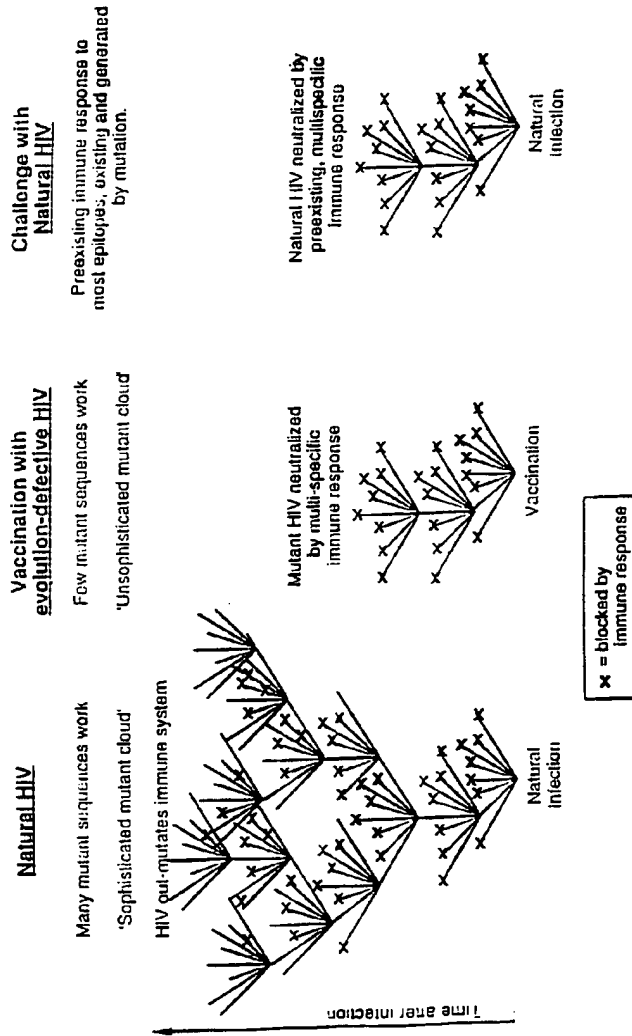


Fig. 16 A

Arg

<u>AGA</u>	<u>CGT</u>	<u>AGA</u>	<u>CGC</u>	<u>AGA</u>	<u>CGA</u>	<u>AGA</u>	<u>CGG</u>
TGA .	AGT S	TGA .	AGC S	TGA .	AGA S	TGA .	AGG S
CGA R	GGT G	CGA R	GGC G	CGA R	GGA G	CGA R	GGG G
GGA G	TGT <u>C</u>	GGA G	TGC <u>C</u>	GGA G	TGA .	GGA G	TGG <u>W</u>
AAA <u>K</u>	CAT <u>H</u>	AAA <u>K</u>	CAC <u>H</u>	AAA <u>K</u>	CAA <u>Q</u>	AAA <u>K</u>	CAG <u>Q</u>
ACA <u>T</u>	CCT <u>P</u>	ACA <u>T</u>	CCC <u>P</u>	ACA <u>T</u>	CCA <u>P</u>	ACA <u>T</u>	CCG <u>P</u>
ATA <u>I</u>	CTT <u>L</u>	ATA <u>I</u>	CTC <u>L</u>	ATA <u>I</u>	CTA <u>L</u>	ATA <u>I</u>	CTG <u>L</u>
AGC S	CGA R	AGC S	CGA R	AGC S	CGC R	AGC S	CGA R
AGT S	CGC R	AGT S	CGT R	AGT S	CGT R	AGT S	CGT R
AGG R	CGG R	AGG R	CGG R	AGG R	CGG R	AGG R	CGC R

<u>AGG</u>	<u>CGT</u>	<u>AGG</u>	<u>CGC</u>	<u>AGG</u>	<u>CGA</u>	<u>AGG</u>	<u>CGG</u>
TGG <u>W</u>	AGT S	TGG <u>W</u>	AGC S	TGG <u>W</u>	AGA S	TGG W	AGG S
CGG R	GGT G	CGG R	GGC G	CGG R	GGA G	CGG R	GGG G
GGG G	TGT <u>C</u>	GGG G	TGC <u>C</u>	GGG G	TGA .	GGG G	TGG W
AAG <u>K</u>	CAT <u>H</u>	AAG <u>K</u>	CAC <u>H</u>	AAG <u>K</u>	CAA <u>Q</u>	AAG <u>K</u>	CAG <u>Q</u>
ACG <u>T</u>	CCT <u>P</u>	ACG <u>T</u>	CCC <u>P</u>	ACG <u>T</u>	CCA <u>P</u>	ACG <u>T</u>	CCG <u>P</u>
ATG <u>M</u>	CTT <u>L</u>	ATG <u>M</u>	CTC <u>L</u>	ATG <u>M</u>	CTA <u>L</u>	ATG <u>M</u>	CTG <u>L</u>
AGA R	CGA R	AGA R	CGA R	AGA R	CGC R	AGA R	CGA R
AGC S	CGC R	AGC S	CGT R	AGC S	CGT R	AGC S	CGT R
AGT S	CGG R	AGT S	CGG R	AGT S	CGG R	AGT S	CGC R

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Fig. 16 B

Ser

<b>AGT</b>	<b>TCT</b>	<b>AGT</b>	<b>TCG</b>	<b>AGT</b>	<b>TCA</b>	<b>AGT</b>	<b>TCG</b>
CGT <b>R</b>	ACT T	CGT <b>R</b>	ACC T	CGT <b>R</b>	ACA T	CGT <b>R</b>	ACG T
GGT <b>G</b>	CCT <b>P</b>	GGT <b>G</b>	CCC <b>P</b>	GGT <b>G</b>	CCA <b>P</b>	GGT <b>G</b>	CCG <b>P</b>
TGT C	GCT <b>A</b>	TGT C	GCC <b>A</b>	TGT <b>C</b>	GCA <b>A</b>	TGT <b>C</b>	GCG <b>A</b>
AAT <b>N</b>	TAT <b>Y</b>	AAT <b>N</b>	TAC <b>Y</b>	AAT <b>N</b>	TAA .	AAT <b>N</b>	TAG .
ACT T	TGT C	ACT T	TGC C	ACT T	TGA .	ACT T	TGG <b>W</b>
ATT <b>I</b>	TTT <b>F</b>	ATT <b>I</b>	TTC <b>F</b>	ATT <b>I</b>	TTA <b>L</b>	ATT <b>I</b>	TTG <b>L</b>
AGA <b>R</b>	TCA S	AGA <b>R</b>	TCA S	AGA <b>R</b>	TCC S	AGA <b>R</b>	TCA S
AGG <b>R</b>	TCG S	AGG <b>R</b>	TCG S	AGG <b>R</b>	TCG S	AGG <b>R</b>	TCC S
AGC S	TCC S	AGC S	TCT S	AGC S	TCT S	AGC S	TCT S

<b>AGC</b>	<b>TCT</b>	<b>AGC</b>	<b>TCG</b>	<b>AGC</b>	<b>TCA</b>	<b>AGC</b>	<b>TCG</b>
CGC <b>R</b>	ACT T	CGC <b>R</b>	ACC T	CGC <b>R</b>	ACA T	CGC <b>R</b>	ACG T
GGC <b>G</b>	CCT <b>P</b>	GGC <b>G</b>	CCC <b>P</b>	GGC <b>G</b>	CCA <b>P</b>	GGC <b>G</b>	CCG <b>P</b>
TGC C	GCT <b>A</b>	TGC C	GCC <b>A</b>	TGC <b>C</b>	GCA <b>A</b>	TGC <b>C</b>	GCG <b>A</b>
AAC <b>N</b>	TAT <b>Y</b>	AAC <b>N</b>	TAC <b>Y</b>	AAC <b>N</b>	TAA .	AAC <b>N</b>	TAG .
ACC T	TGT C	ACC T	TGC C	ACC T	TGA .	ACC T	TGG <b>W</b>
ATC <b>I</b>	TTT <b>F</b>	ATC <b>I</b>	TTC <b>F</b>	ATC <b>I</b>	TTA <b>L</b>	ATC <b>I</b>	TTG <b>L</b>
AGA <b>R</b>	TCA S	AGA <b>R</b>	TCA S	AGA <b>R</b>	TCC S	AGA <b>R</b>	TCA S
AGG <b>R</b>	TCG S	AGG <b>R</b>	TCG S	AGG <b>R</b>	TCG S	AGG <b>R</b>	TCC S
AGT S	TCC S	AGT S	TCT S	AGT S	TCT S	AGT S	TCT S

Fig. 16

Leu

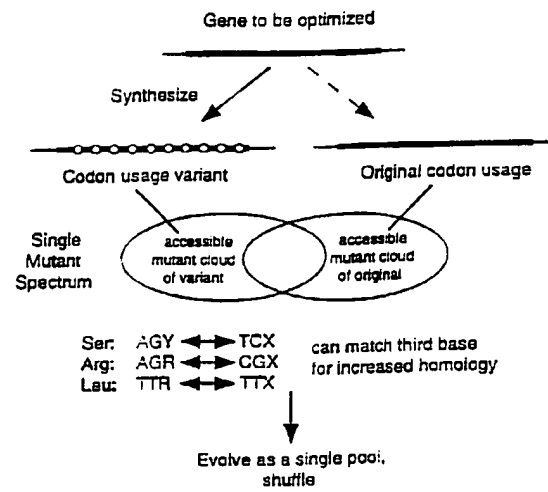
<u>TTA</u>	<u>CTT</u>	<u>TTA</u>	<u>CTC</u>	<u>TTA</u>	<u>CTA</u>	<u>TTA</u>	<u>CTG</u>
ATA I	ATT I	ATA I	ATC I	ATA I	ATA I	ATA I	ATG M
CTA L	GTT V	CTA L	GTC V	CTA L	GTA V	CTA L	GTG V
GTA V	TTT F	GTA V	TTC F	GTA V	TTA F	GTA V	TTG L
TAA .	CAT <b>H</b>	TAA .	CAC <b>H</b>	TAA .	CAA <b>H</b>	TAA .	CAG <b>Q</b>
TCA <b>S</b>	CCT <b>P</b>	TCA <b>S</b>	CCC <b>P</b>	TCA <b>S</b>	CCA <b>P</b>	TCA <b>S</b>	CCG <b>P</b>
TGA .	CGT <b>R</b>	TGA .	CGC <b>R</b>	TGA .	CGA <b>R</b>	TGA .	CGG <b>R</b>
TTG L	CTA L	TTG L	CTT L	TTG L	CTT L	TTG L	CTT L
TTC F	CTC L	TTC F	CTA L	TTC F	CTC L	TTC <b>F</b>	CTC L
TTT F	CTG L	TTT F	CTG L	TTT F	CTG L	TTT <b>F</b>	CTA L

<u>TTG</u>	<u>CTT</u>	<u>TTG</u>	<u>CTC</u>	<u>TTG</u>	<u>CTA</u>	<u>TTG</u>	<u>CTG</u>
ATG <b>M</b>	ATT I	ATG <b>M</b>	ATC I	ATG <b>M</b>	ATA I	ATG M	ATG M
CTG L	GTT V	CTG L	GTC V	CTG L	GTA V	CTG L	GTG V
GTG V	TTT F	GTG V	TTC F	GTG V	TTA F	GTG V	TTG L
TAG .	CAT <b>H</b>	TAG .	CAC <b>H</b>	TAG .	CAA <b>H</b>	TAG .	CAG <b>Q</b>
TCG <b>S</b>	CCT <b>P</b>	TCG <b>S</b>	CCC <b>P</b>	TCG <b>S</b>	CCA <b>P</b>	TCG <b>S</b>	CCG <b>P</b>
TGG <b>W</b>	CGT <b>R</b>	TGG <b>W</b>	CGC <b>R</b>	TGG <b>W</b>	CGA <b>R</b>	TGG <b>W</b>	CGG <b>R</b>
TTA I	CTA L	TTA I	CTT L	TTA I	CTT L	TTA <b>I</b>	CTT L
TTC F	CTC L	TTC F	CTA L	TTC F	CTC L	TTC <b>F</b>	CTC L
TTT F	CTG L	TTT F	CTG L	TTT F	CTG L	TTT <b>F</b>	CTA L

Fig. 17

### Protein Evolution with Expanded Mutation Spectra



Best synthesis method: all possible codons based on codon synthesis

## Fig 18A

## Altering the evolution potential of sequences by codon modification

HIV envelope protein (p57 amino acids)

Potential mutant resulting from wildtype sequence versus codon modified sequence in arg, leu and serine codons:

[illegible]

**Codon-modified**





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## 188 FINAL ALIGNMENT

Saturday, August 17, 1 8 2:03 PM

Lipman-Pearson Protein Alignment

kTuplet: 2; Gap Penalty: 4; Gap Length Penalty: 12

Seq1(1&gt;509)

Seq2(1&gt;509)

Similarity

Gap

Gap

Consensus

new env pro

old env protein

Index

Number

Length

Length

(1&gt;509)

(1&gt;509)

100.0

0

0

509

10 20 30 40 50 60 70  
 YQYVTVFYGVPAWKNAIPLFCATKNRDTWGTTCCLPONDDYSELAINVTEAFDAWONTVTEQAIEDVWN  
 YQYVTVFYGVPAWKNAIPLFCATKNRDTWGTTCCLPONDDYSELAINVTEAFDAWONTVTEQAIEDVWN  
 YQYVTVFYGVPAWKNAIPLFCATKNRDTWGTTCCLPONDDYSELAINVTEAFDAWONTVTEQAIEDVWN  
 10 20 30 40 50 60 70  
 80 90 100 110 120 130 140  
 LFETSIKPCVKLTPLCIAMRCNKTEDEWGLTGNAGTTTATTTATPSVAENVINESNPCIKNNSCAGL  
 LFETSIKPCVKLTPLCIAMRCNKTEDEWGLTGNAGTTTATTTATPSVAENVINESNPCIKNNSCAGL  
 LFETSIKPCVKLTPLCIAMRCNKTEDEWGLTGNAGTTTATTTATPSVAENVINESNPCIKNNSCAGL  
 80 90 100 110 120 130 140  
 150 160 170 180 190 200 210  
 EQEPHIGCKFNMGTGLNRDCKKEYNETWYSRDLICEGSANESESKCYMHHCNTSVIQESCKHYWDAIRFR  
 EQEPHIGCKFNMGTGLNRDCKKEYNETWYSRDLICEGSANESESKCYMHHCNTSVIQESCKHYWDAIRFR  
 EQEPHIGCKFNMGTGLNRDCKKEYNETWYSRDLICEGSANESESKCYMHHCNTSVIQESCKHYWDAIRFR  
 150 160 170 180 190 200 210  
 220 230 240 250 260 270 280  
 YCAPPGYALLRCNDSNYLGFAPNCSKYVVSCTRMHMETCTSTWFGFNGTRAENRTYIYWHGKSNRTIISL  
 YCAPPGYALLRCNDSNYLGFAPNCSKYVVSCTRMHMETCTSTWFGFNGTRAENRTYIYWHGKSNRTIISL  
 YCAPPGYALLRCNDSNYLGFAPNCSKYVVSCTRMHMETCTSTWFGFNGTRAENRTYIYWHGKSNRTIISL  
 220 230 240 250 260 270 280  
 290 300 310 320 330 340 350  
 NKYYNLTMRCPGPKNTVLPVTIMSGLVFHSQINERPKQAWCFEGSWKKAIOEVKETLVKHPRYTGTN  
 NKYYNLTMRCPGPKNTVLPVTIMSGLVFHSQINERPKQAWCFEGSWKKAIOEVKETLVKHPRYTGTN  
 NKYYNLTMRCPGPKNTVLPVTIMSGLVFHSQINERPKQAWCFEGSWKKAIOEVKETLVKHPRYTGTN  
 290 300 310 320 330 340 350  
 360 370 380 390 400 410 420  
 DTRKINLTAPAGGPEVTFMWTNCRGEFLYCKMNWFLNWVEDRQKGGRWKQNRKEQOKXNYYPCHIRQ  
 DTRKINLTAPAGGPEVTFMWTNCRGEFLYCKMNWFLNWVEDRQKGGRWKQNRKEQOKXNYYPCHIRQ  
 DTRKINLTAPAGGPEVTFMWTNCRGEFLYCKMNWFLNWVEDRQKGGRWKQNRKEQOKXNYYPCHIRQ  
 360 370 380 390 400 410 420  
 430 440 450 460 470 480 490  
 IINTWHKVGKNVYLPREGDLTCNSTYTSLIAEIDWINSNETNITHSAEVAELRYLELGDYKLEITPIG  
 IINTWHKVGKNVYLPREGDLTCNSTYTSLIAEIDWINSNETNITHSAEVAELRYLELGDYKLEITPIG  
 IINTWHKVGKNVYLPREGDLTCNSTYTSLIAEIDWINSNETNITHSAEVAELRYLELGDYKLEITPIG  
 430 440 450 460 470 480 490  
 500  
 LAPTSVRRYTTTGASRNKR  
 LAPTSVRRYTTTGASRNKR  
 LAPTSVRRYTTTGASRNKR  
 500

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185      186  
DNA HOMOL 3Y

Page 1

Saturday, August 17, 1999 4:39 PM

Wilbur-Lipman DNA Alignment

Ktuple: 3; Gap Penalty: 20; Window: 20

Seq1(1&gt;1527)

Seq2(1&gt;1527)

new env

old env

Similarity Index	Gap Number	Gap Length	Consensus Length
(61.7)	0	0	1445

```

(10>1454)      (10>1454)
#10      #20      #30      #40      #50      #60
GTBACBGTBTTTACGGAGTTCTGCTTGGAAAAACGCHACBATHCCOTTRTTTTCGGCT
GT AC GT TT TA GG GT CC GC TGGAA AA GC AC AT CC T TT TG GC
GTAACAGTATTCTATGGTGTACCGCATGGAAGAATGCGACAATTCCCTCTTCTGTGCA
#10      #20      #30      #40      #50      #60
#70      #80      #90      #100     #110     #120
ACAAAAAACCGHGTACVTTGGGGSACBACBAGTGTCTCCCTGACAACGACGACTATAGY
AC AA AA G GA AC TGGGG AC AC CA TG T CC GA AA GA GA TA
ACCAAGAATAGGGACACTTGGGGAAACAACACAATGCTTGCAGATAATGATGATTACTCA
#70      #80      #90      #100     #110     #120
#130     #140     #150     #160     #170     #180
GAGCTHGCBATAAACGTAACCTGAAGCATTGACGCVTGGGACAACACBGTDACGTAGCAG
GA T GC AT AA GT AC GA GC TT GA GC TGGGA AA AC GT AC GA CA
GAATTGGCAAATCAATGTACAGAGGGCTTTTGATGCTTGGGATAATACAGTCACAGAAACA
#130     #140     #150     #160     #170     #180
#190     #200     #210     #220     #230     #240
GCTATTGAAGACGTHTGGAAATTTTCGAGACTAGTATAAAACCATGCGTTAAGTTTRACO
GC AT GA GA GT TGGAA T TT GA AC AT AA CC TG GT AA T AC
GCAATAGAGGATGTGTGGAACTCTTTGAAACATCCATTAAGCCCTGTGTAAACTCACC
#190     #200     #210     #220     #230     #240
#250     #260     #270     #280     #290     #300
CCBTGTGCTGCTATGAGGTGCAACAGACVGAACBGACCGHTGGGGVCTCACTGGT
CC T TG AT GC ATGAG TG AA AA AC GA AC GA G TGGGG T AC GG
CCACTATGTATAGCAATGAGATGTAATAAACTGAGACAGATAGGTGGGGTTTGACAGGA
#250     #260     #270     #280     #290     #300
#310     #320     #330     #340     #350     #360
AATGCTGGTACBACBACBACBACBACBACBACBACBACBACBACBACBACBACBACBAC
AA GC GG AC AC AC AC GC AT AC AC AC GC AC CC GT GC GA AA
AACGCAGGGACAACAACAACAGCAATAACAACAACAGCAACACCAAGTGTAGCAGAAAAAT
#310     #320     #330     #340     #350     #360
#370     #380     #390     #400     #410     #420
GTATYAACGAGTCAAACCCATGTATTAAGAACAACCTVTGCGCGGGCTYAGCAAGAA
GT AT AA GA AA CC TG AT AA AA AA TG GC GG T GA CA GA
GTTATAAATGAAGTAATCCTTGCATAAAAAATAATAGTTGTCCAGGCTTGGAAACAGGAG
#370     #380     #390     #400     #410     #420
#430     #440     #450     #460     #470     #480
CCDATGATTGGVTCGAAGTTCAATATGACBGGCTTAACCGCCATAAGAAAAAGGAGTAC
CC ATGAT GG TG AA TT AA ATGAC GG T AA G GA AA AA AA GA TA
CCCATGATAGGTTGTAAATTTAACATGACAGGGTTAAATAGGACAAAAAAGAAAGATAT
#430     #440     #450     #460     #470     #480
#490     #500     #510     #520     #530     #540
AACGAGACBTGGTACAGTCGTGACCTTATATGCGATCAAAGYGCHACGAGTCAGATCC
AA GA AC TGGTA G GA T AT TG GA CA GC AA GA GA
AATGAACATGGTATTCAAGAGATTTAATCTGTGAGCAGTCAGCGAATGAAAGTGAGAGT
#490     #500     #510     #520     #530     #540
#550     #560     #570     #580     #590     #600
AAGTGCTATATGACCACATGCAATACDTCVGTAAATACAGGAGAGTTGCGATAAACACTAC
AA TG TA ATGCA CA TG AA AC GT AT CA GA TG GA AA CA TA
AAATGTTACATGCATCATTTGTAACACCAAGTGTATTCAAGAATCTGTGACAAGCATTTAT
#550     #560     #570     #580     #590     #600

```

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Fig. 18C

Page 2

Saturday, August 17, 1996 4:39 PM  
 Wilbur-Lipman DNA Alignment  
 Ktuple: 3; Gap Penalty: 20; Window: 20  
 Seq1(1>1527) Seq2(1>1527)  
 new env.DNA old env.pro

		Similarity	Gap	Gap	Consensus
		Index	Number	Length	Length
(10>1454)	(10>1454)	61.7	0	0	1445
#610	#620	#630	#640	#650	#660
TGGGACGCVATACGTTTCCGCTATTGCGCTCCHCCBGGVTACGCVCTHTTTCGCTGCAAC					
TGGGA GC AT G TT G TA TG GC CC CC GG TA GC T T G TG AA					
TGGGATGCTATTAGATTAGATACTGTGCACCGCCAGGTTATGCTTTGCTTAGGTGTAAT					
#610	#620	#630	#640	#650	#660
#670	#680	#690	#700	#710	#720
GACAGTAACCTACCTBGGDTTCGCVCCVAATTGCAGYAAAGTTGCTGAAGCAGYGTACB					
GA AA TA T GG TT GC CC AA TG AA GT GT GT TG AC					
GATTCAAATTATTAGGCTTTGCTCCTAACTGTTCTAAGGTAGTGGTTTCTTCATGCACA					
#670	#680	#690	#700	#710	#720
#730	#740	#750	#760	#770	#780
CGBATGATGGAAACCCAGACGAGCACATGLTTCGGDTTAAACGGYACVCGHGETGAGAAC					
G ATGATGGA AC CA AC AC TGGTT GG TT AA GG AC G GC GA AA					
AGAATGATGGAGACGCAAAACCTCTACTTGGTTTGGCTCAATGGTACTAGGGCAGAAAAAT					
#730	#740	#750	#760	#770	#780
#790	#800	#810	#820	#830	#840
CGTACTTATATHTACTGGCACGGDAAGTCVAACCGTACGATTATATCGCTCAACAAATAC					
G AC TA AT TA TGGA GG AA AA G AC AT AT T AA AA TA					
AGAACATACATTATTGGCATGGCAAAAGTAATAGAACCAATAATTAGCTTGAATAAGTAT					
#790	#800	#810	#820	#830	#840
#850	#860	#870	#880	#890	#900
TACAACTTYACTATGCGTTGCCGTGCCCTGGBAACAAAACBGTCTBCEBGTAAACGATA					
TA AA T AC ATG G TG G G CC GG AA AA AC GT T CC GT AC AT					
TATAATCTAACAAATGAGATGTAGAAGACCAGGAAATAAGACAGTTTACCAGTCACCATT					
#850	#860	#870	#880	#890	#900
#910	#920	#930	#940	#950	#960
ATGACTGGCCTHGTDTTTCACAGYCAACCCATTAACGAACGTCCTAAGCAAGCDTGGTGT					
ATG C GG T GT TT CA CA CC AT AA GA G CC AA CA GC TGGTG					
ATGTCAGGGTTGGTCTTCCATTCCGAGCCCATAAATGAGAGACCAAAACAGGCTTGGTGC					
#910	#920	#930	#940	#950	#960
#970	#980	#990	#1000	#1010	#1020
TGGTTCGAGGGTTCTGTGGAAGAAAGCGATACAAGAGGTHAAAGAGACDCTHGTGAAGCAC					
TGGTT GA GG TGGAA AA GC AT CA GA GT AA GA AC T GT AA CA					
TGGTTTGAAGGAAGCTGGAAAAGGCCATCCAGGAAGTGAAGGAAACCTTGGTCAACAT					
#970	#980	#990	#1000	#1010	#1020
#1030	#1040	#1050	#1060	#1070	#1080
CCGCGCTACACHGGGACVAACGACACVCGHAAGATAAACTTGACTGCACCTGCBGGGBGB					
CC G TA AC GG AC AA GA AC G AA AT AA T AC GC CC GC GG GG					
CCAGGTATACGGGAACATAATGATACTAGGAAAATTAATCTAACAGCTCCAGCAGGAGGA					
#1030	#1040	#1050	#1060	#1070	#1080
#1090	#1100	#1110	#1120	#1130	#1140
GACCCSGAGGTGACATTTCATGTGGACTAACTGCAGRGGGAGTTTCTBTACTGTAAGATG					
GA CC GA GT AC TT ATGTGGAC AA TG G GG GA TT T TA TG AA ATG					
GATCCAGAACTCACTTTTATGTGGACAAATTGTCGAGGAGAATTCTTATATTGCAAAATG					
#1090	#1100	#1110	#1120	#1130	#1140
#1150	#1160	#1170	#1180	#1190	#1200
AACCTGTTCTTAACTGGGTBGAAGATCGGATCAGAAAGGAGGGCGTTGGAAGCAGCAG					
AA TGGTT - AA TGGGT GA GA G GA CA AA GG GG G TGGAA CA CA					
AATTGTTTCTTAATTGGGTAGAGGACAGAGACCAAAAGGGTGGCAGATGCAACAACAA					
#1150	#1160	#1170	#1180	#1190	#1200

Fig 18C

Page 3

Saturday, August 17, 1996, 4:39 PM

Wilbur-Lipman DNA Alignment

Ktuple: 3; Gap Penalty: 20; Window: 20

Seq1(1&gt;1527)

Seq2(1&gt;1527)

new env DNA

old env.pro

Seq1(1>1527)	Seq2(1>1527)	Similarity Index	Gap Number	Gap Length	Consensus Length
(10>1454)	(10>1454)	61.7	0	0	1445
#1210 #1220 #1230 #1240 #1250 #1260 AACCGHAAGGAACAGCAAAAAAGAACTACGTHCCBTGCCACATMCGBCAGATTATAAAT AA G AA GA CA CA AA AA TA GT CC TG CA AT G CA AT AT AA AATAGGAAAGAGCAACAGAAAGAAAAATTATGTGCCATGTCATATTAGACAAATAATCAAC #1210 #1220 #1230 #1240 #1250 #1260 #1270 #1280 #1290 #1300 #1310 #1320 ACCTGGCATAAGGTSGGCAAGAACGTBTACCTCCCAACCCCGGAGGGTGATTTRACBTGT AC TGGCA AA GT GG AA AA GT TA T CC CC G GA GG GA T AC TG ACCTGGCACAAAGTAGGCAAAATGTATATTTGCCCTCCTAGGGAAGGAGACCTGACATGC #1270 #1280 #1290 #1300 #1310 #1320 #1330 #1340 #1350 #1360 #1370 #1380 AACAGYACVGTACATCATTGATTCTGAAATYGAATGGAATWAACCTDAACGAAACGAAC AA AC GT AC T AT GC GA AT GA TGGAT AA AA GA AC AA AATTCACCTGTAAC TAGTCTCATAGCAGAGATAGATTGGATCAATAGCAATGAGACCAAT #1330 #1340 #1350 #1360 #1370 #1380 #1390 #1400 #1410 #1420 #1430 #1440 ATAACCATGTCVGCBAAGTHGCBGAGTYYACAGGCTCGAACTCGGTGACTATAAGCTB AT AC ATG GC GA GT GC GA T TA G T GA T GG GA TA AA T ATCACCATGAGTGCAGAGGTGGCAGAACTGTATCGATTGGAGTTGGGAGATTACAAATTA #1390 #1400 #1410 #1420 #1430 #1440 #1450 ATYGA AT GA ATAGA #1450					

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Saturday, August 17, 1996 1:31 PM  
1.8 kb Env mutagenized  
Created: Friday, August 16, 1996 9:39 AM

F5

18d

Page 1

MUTAGENIZED GENE  
IN CONTEXT

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      10      20      30      40
.....
aacctccgaaggaagactaaggctaatccatttcctgcatca 40
aaccaagtaagatagggatgtctctgggaatcagctgcttat 80
cgcgcctcttgetagtaagtggttttcgagatttgggtgtv 120
CAGTAcGTbACbGTbTTtTAcGGaGTTcCTGCTTGGAAdA 160
AcGGChACbATmCCdtttrttTGcGCTACaAAaAAccghGA 200
      210      220      230      240
.....
tACvTGGGGbACbACbCAGTgtctcCTGACaACGAcGAc 240
TAtagyGAGctnGCbATcAAcGTaACTGAaGCATTcGAcG 280
CvTGGGACaAaACbGTdACTGAGCAGGCTATTGAaGAcGT 320
hTGGAAttttrTTcGAGAcagTATaAAcCCaTGcGTTAAg 360
tttACdCCbtttgTGCATTGCTATGAGgTGcAAcAAGAcvG 400
      410      420      430      440
.....
AaACbGAccgnTGGGGvctcACTGGtAAtGCTGGtACbAC 440
bACbACbGCbATyACbACtACcGCTACbCCttcvGTtGcb 480
GAGAAcGTyATyAAcGAGtcaAAcCCaTGtATTAAgAAcA 520
ActevTGcGCBGGdctyGAGCAaGAaCCdATGATtGGvTG 560
cAAgTTcAAtATGACbGGccttAaccgcGAtAAgAAaAAg 600
      610      620      630      640
.....
GAGTAcAAcGAGAcBTGGTAcagtcgtGAccttATaTGcG 640
AaCAacgyGChAAcGAGtcaGAatccAAgTGcTAtATGCA 680
cCAcTGcAAATAcDtevGTaATaCAGGAgagtTGcGATAaA 720
CAcTAcTGGGACGCvATacgtTTccgcTAtTGcGCTcChC 760
CbGGvTAcGCVcthtttregeTGcAAcGAcagtAAcTAcct 800
      810      820      830      840
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bGGdTTCGcVCCvAAATGcagyaAaGTTGTcGTaagcagy 840
TGtACbcbATGATGGAaACcCAGAcgagcACaTGGTTcG 880
GdTTtAAcGGvACvcghGCTGAGaAaccgtACTtATAtmTA 920
cTGGCAGGGdAAgtcvaAaccgtACgATTAtatcgtcAAC 960
AAaTAcTAcAAccttgACtATGcgtTGccgtcgcCTGGbA 1000
      1010      1020      1030      1040
.....
AcAAaACbGTvctbCCbGTaACgATaATGagtGGcctnGT 1040
dTTtCAcagycAAcCdATTtAAcGAacgtCCtAAgCAaGcd 1080
TGGTgtTGGTTcGAGGGttcgTGGAAgAAaGCgATaCAaG 1120
AgGThAAaGAGAcDctnGTgAAgCAcCCgcgcTAcACHGG 1160
bACvAAcGAcAcvcgnAAgATaAAccttgACTGcAcCTGcb 1200

```

Figure 19

## ENV Top oligos

1T CTAGTAAAGGTTTATGAGATTGTTGT  
2T TTACGGAGTTCCTGCTTGGAAAACGGTACCATTCCTTTTTCGGCTACAAAAAAGG  
3T AGTGTCTCCCTGACAAAGAGCTATAGTGCCTTCCATAAAGTAACTGAGCTTGG  
4T AGTGGCTCCCTATTTAGAGGTTTGGATTCTTTCAGAGCTTATATAAACCATCGGT  
5T TCGTATCGGGTGCACACACACGAAACCGAGGTTGGGVTCTACCTGGTATCGCTGAC  
6T CTTACGGCTACCGCTTCTGTTCCGAGAGGTTATTAAGAGTCAAAACCTGTATATAGA  
7T GCGCAGAGAGCGGTGTTGGVTGCAAGTTCAATATGACCGGCTTAAACCGGTATAGAA  
8T GTACAGTGTGACCTTATATCGGACAAAGTGGTAAAGTACAGATCCATGTTGCTATAT  
9T TATACAGGAGGTTTCGGTAAACACTACTCGGAGGCTATAGCTTCCCTATTCGGCTC  
10T CGCTGCAAGGACAGTACTTCCCTTCCGCTTCCGCTTCCGCTTCCGCTTCCGCTTCCGCT  
11T AAACCGAGAGGACATGTTGGTCTTAAAGGACVCGGCTGACAGACCGTACTTATAT  
12T AACGTGAGTATATATGCTTCAACAAATCTTCACTTCACTTCACTTCACTTCACTTCACT  
13T GTAAAGTAAATGAGTGGCTTCTTTCACAGTCAACCTATTAAGGAGGTTCTTANGAA  
14T TTCTGCGAAGAAAGGATACAGAGGTAAGAGAGTCTTCTTCTTCTTCTTCTTCTTCTTCT  
15T AGTAAACTTCTCTTCACTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCT  
16T TGTAGATGACCTGCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCT  
17T ACAGCAAAAAAGACTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCT  
18T TCCCAACCGCTGCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCT  
19T GAAAGGACATAAGTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCT  
20T ACCTATAGGCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCT

